

PET-MRI in neuroimmunological diseases: the Paris experience with TSP0-PET applied to MS

International OMS workshop

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Paris, France

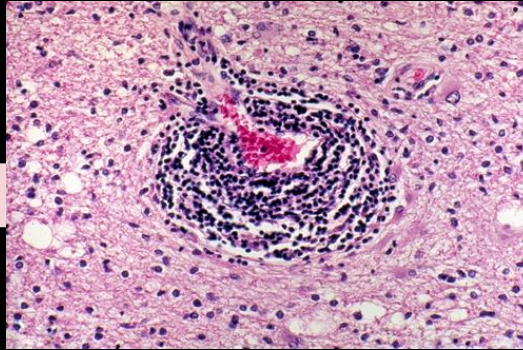
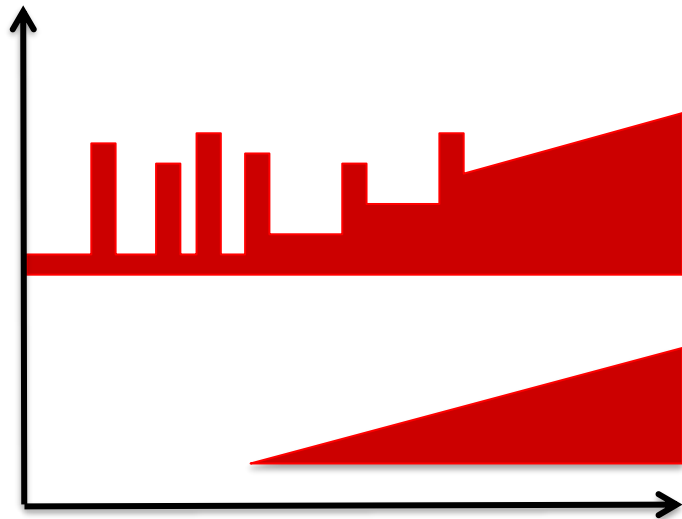


- Why and how using PET to explore neuroinflammation in MS?
- Exploring neuroinflammation in lesions and normal-appearing tissues in MS
- Regionalization of neuroinflammation in the MS brain
- A dysfunction of the Brain/CSF barrier at the choroid plexus level

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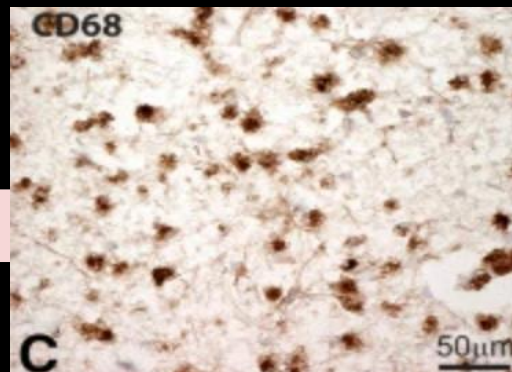
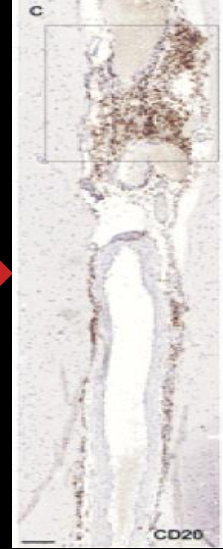
Adaptive and innate immune systems in the pathophysiology of neurodegeneration

Neurodegeneration



Adaptive immune system

- Acute demyelination
- Axonal transection
- Ectopic lymphocytic inflammation



Innate immune system

- Myelin destruction in acute lesions
- Smouldering plaques
- Diffuse damage in normal-appearing tissues

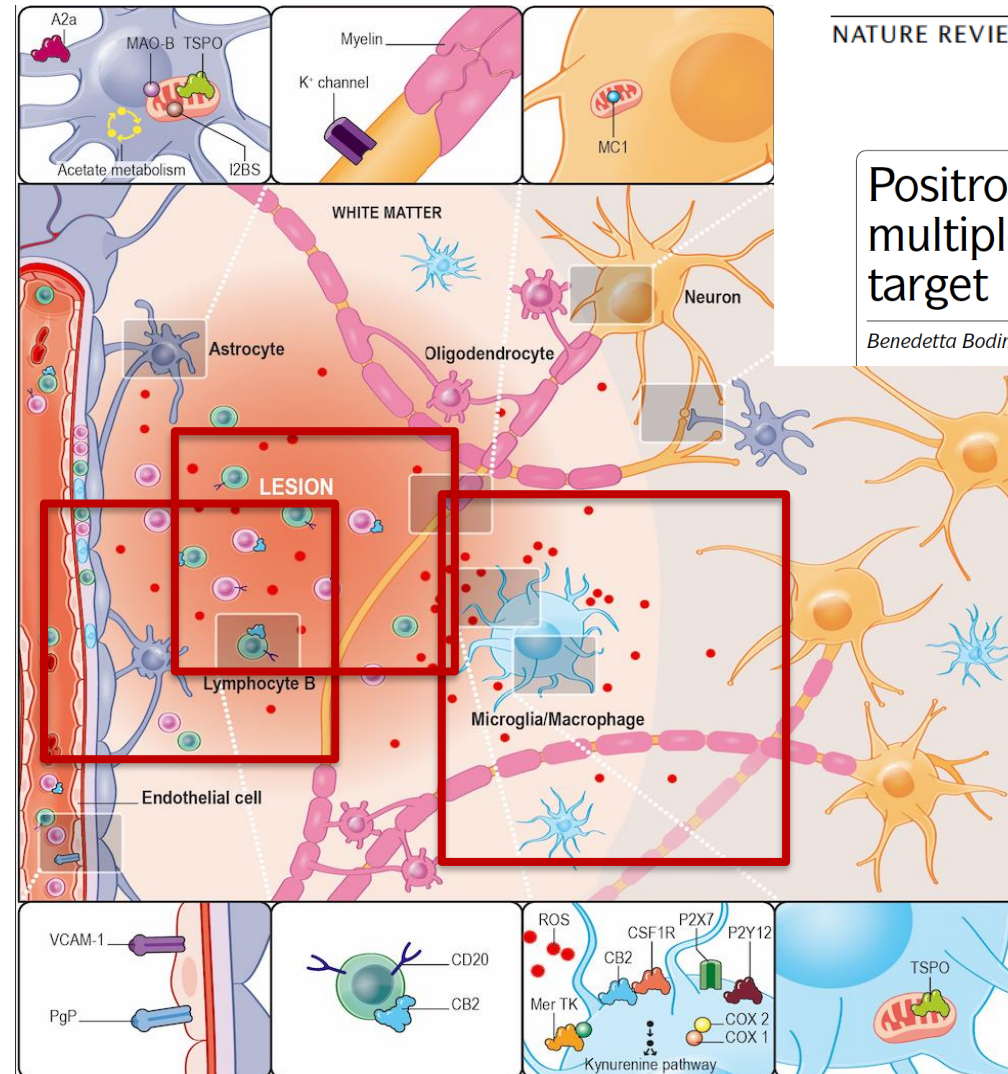


Targeting neuroinflammation with PET

PET



- High specificity for cellular/tissular targets
- Generally low resolution



NATURE REVIEWS | NEUROLOGY

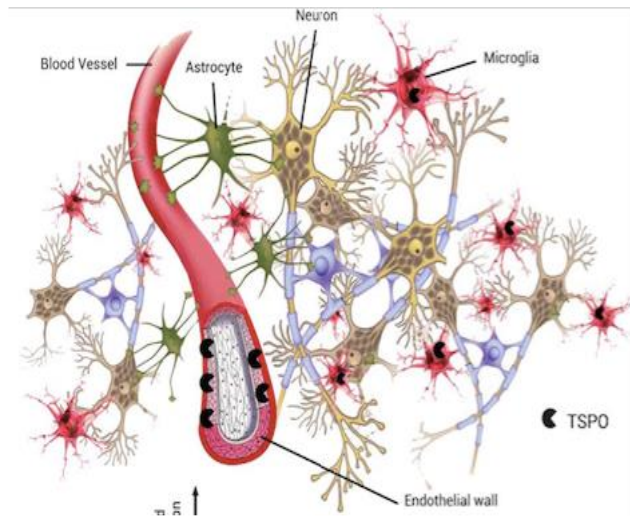
REVIEWS

Check for updates

Positron emission tomography in multiple sclerosis — straight to the target

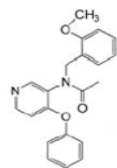
Benedetta Bodini^{1,2}✉, Matteo Tonietto^{1,3}, Laura Airas⁴ and Bruno Stankoff^{1,2}

TSPO

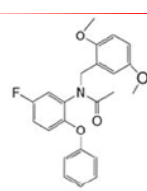


- Macromolecular complex localized in the outer mitochondrial membrane
- Expression mainly driven by innate immune cell activation

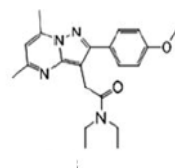
PBR 28



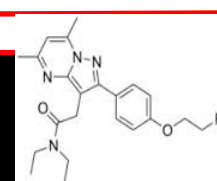
DAA 1106



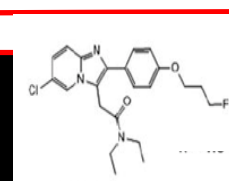
DPA 713



DPA 714



PBR 111



(Benavides, 1993;1988; Le fur 1988)

Improving the signal-to-noise ratio using second-generation TSPO tracers

1st generation

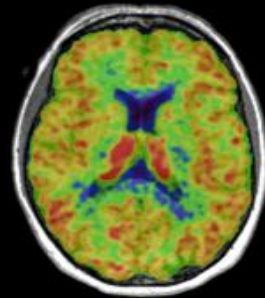
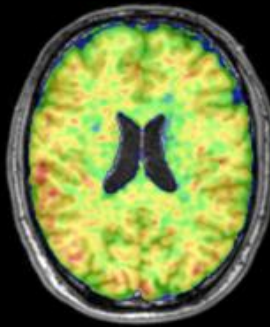
2nd generation

A) ¹¹C-PK11195

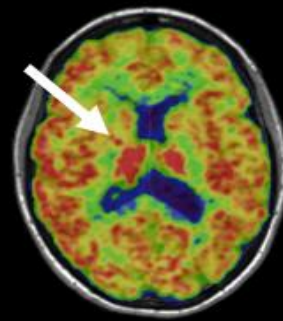
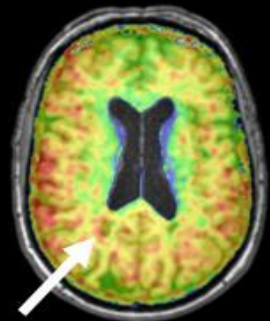
B) ¹⁸F-DPA714

DVR
2.0
1.8
1.6
1.4
1.2
1.0
0.8
0.6
0.4
0.2
0.0

Healthy controls



Patients with MS



- Improved SNR
- Improved affinity

TSPO genetic polymorphism

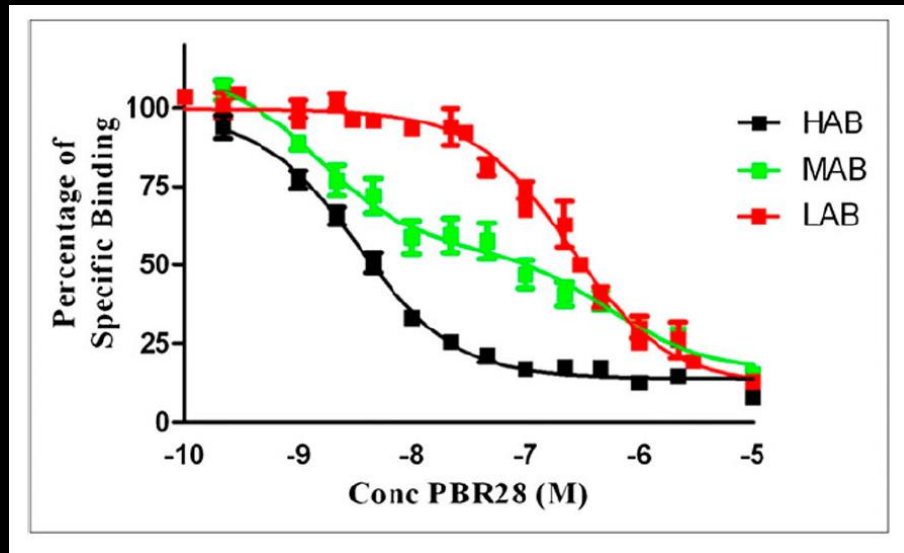
J Nucl Med. 2011 January ; 52(1): 24–32. doi:10.2967/jnumed.110.079459.

Mixed-Affinity Binding in Humans with 18-kDa Translocator Protein Ligands

David R.J. Owen^{1,2}, Roger N. Gunn^{2,3}, Eugenii A. Rabiner^{2,3}, Idriss Bennacef², Masahiro Fujita⁴, William C. Kreisl⁴, Robert B. Innis⁴, Victor W. Pike⁴, Richard Reynolds⁵, Paul M. Matthews^{2,3}, and Christine A. Parker²

An 18-kDa Translocator Protein (TSPO) polymorphism explains differences in binding affinity of the PET radioligand PBR28

David R Owen^{1,2,6}, Astrid J Yeo^{3,6}, Roger N Gunn^{2,4,5}, Kijoung Song³, Graham Wadsworth², Andrew Lewis¹, Chris Rhodes¹, David J Pulford³, Idriss Bennacef², Christine A Parker^{2,4}, Pamela L StJean³, Lon R Cardon³, Vincent E Mooser³, Paul M Matthews^{2,4}, Eugenii A Rabiner^{2,4} and Justin P Rubio³

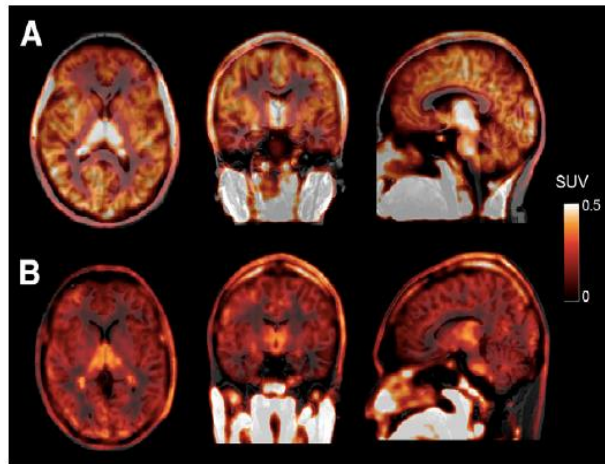


Ligand	LAB	MAB	HAB
PBR28	1	28.2	55.3
PBR06	1	9.2	17.3
DAA1106	1	2.9	4.7
PBR111	1	2.5	4.0
DPA173	1	2.7	4.4
PK11195	1	0.9	0.8

Quantification of ^{18}F -DPA-714

Optimized Quantification of Translocator Protein Radioligand ^{18}F -DPA-714 Uptake in the Brain of Genotyped Healthy Volunteers

Sonia Lavisse^{1,2}, Daniel García-Lorenzo^{3,4}, Marie-Anne Peyronneau⁵, Benedetta Bodini³⁻⁶, Claire Thiriez^{1,2,7}, Bertrand Kuhnast⁵, Claude Comtat⁵, Philippe Remy^{*1,2,7,8}, Bruno Stankoff^{*3,4}, and Michel Bottlaender^{5,9}



Reliable quantification using the 2-TC model
Equilibrium at 60 min
Lavisse et al, J Nuc Med 2015

Original Article **JCBFM**
Journal of Cerebral Blood Flow & Metabolism
2018, Vol. 38(2) 333-346
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DOI: 10.1177/0271678X17692599
journals.sagepub.com/home/jcbfm
SAGE

Validation of an automatic reference region extraction for the quantification of [^{18}F]DPA-714 in dynamic brain PET studies

Daniel García-Lorenzo^{1,*}, Sonia Lavisse^{2,3,*}, Claire Leroy^{4,5}, Catriona Wimberley^{4,5}, Benedetta Bodini¹, Philippe Remy^{2,3,7}, Mattia Veronese⁶, Federico Turkheimer⁶, Bruno Stankoff^{1,8,*} and Michel Bottlaender^{4,5,9,*}

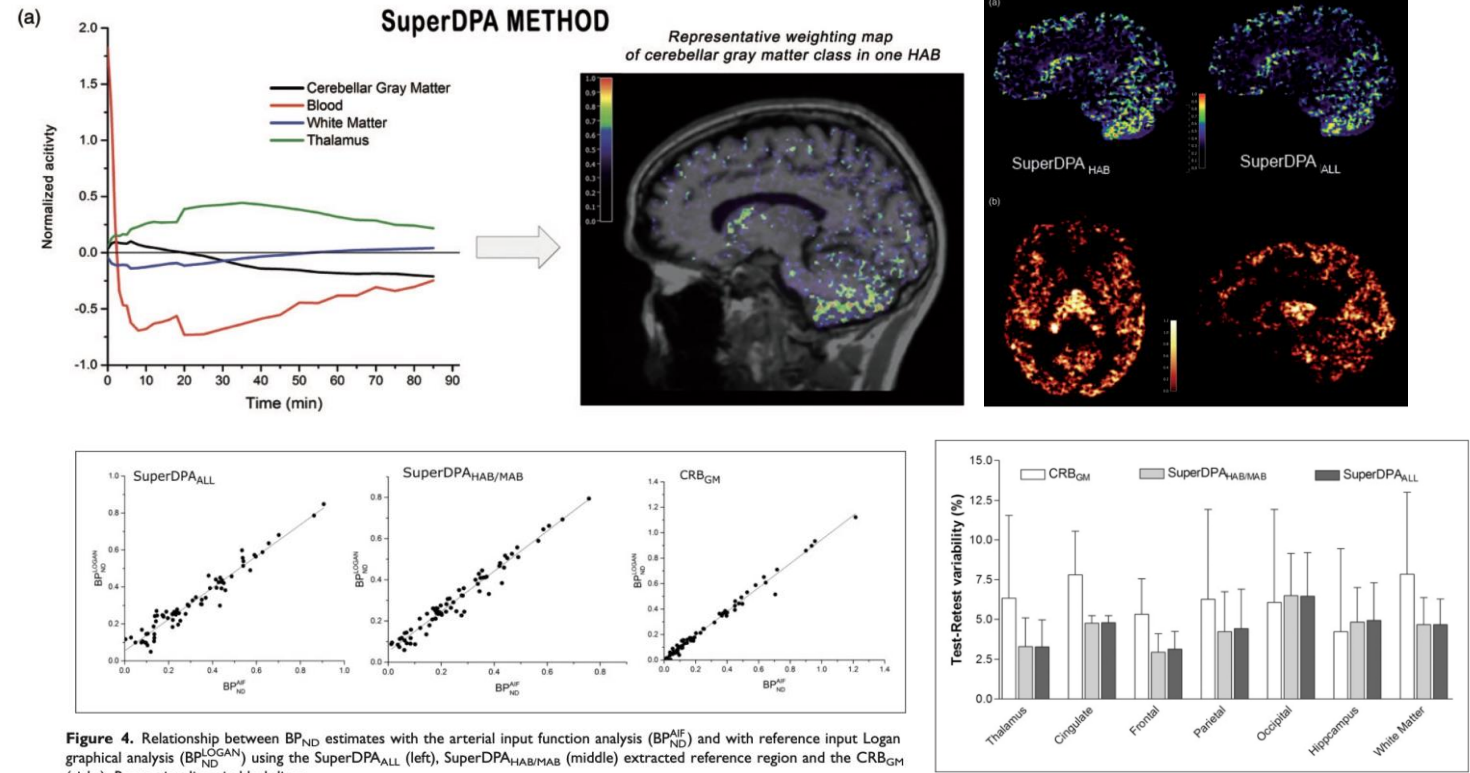


Figure 4. Relationship between BP_{ND} estimates with the arterial input function analysis (BP_{ND}^{AIF}) and with reference input Logan graphical analysis (BP_{ND}^{LOGAN}) using the SuperDPA_{ALL} (left), SuperDPA_{HAB/MAB} (middle) extracted reference region and the CRB_{GM} (right). Regression lines in black lines.

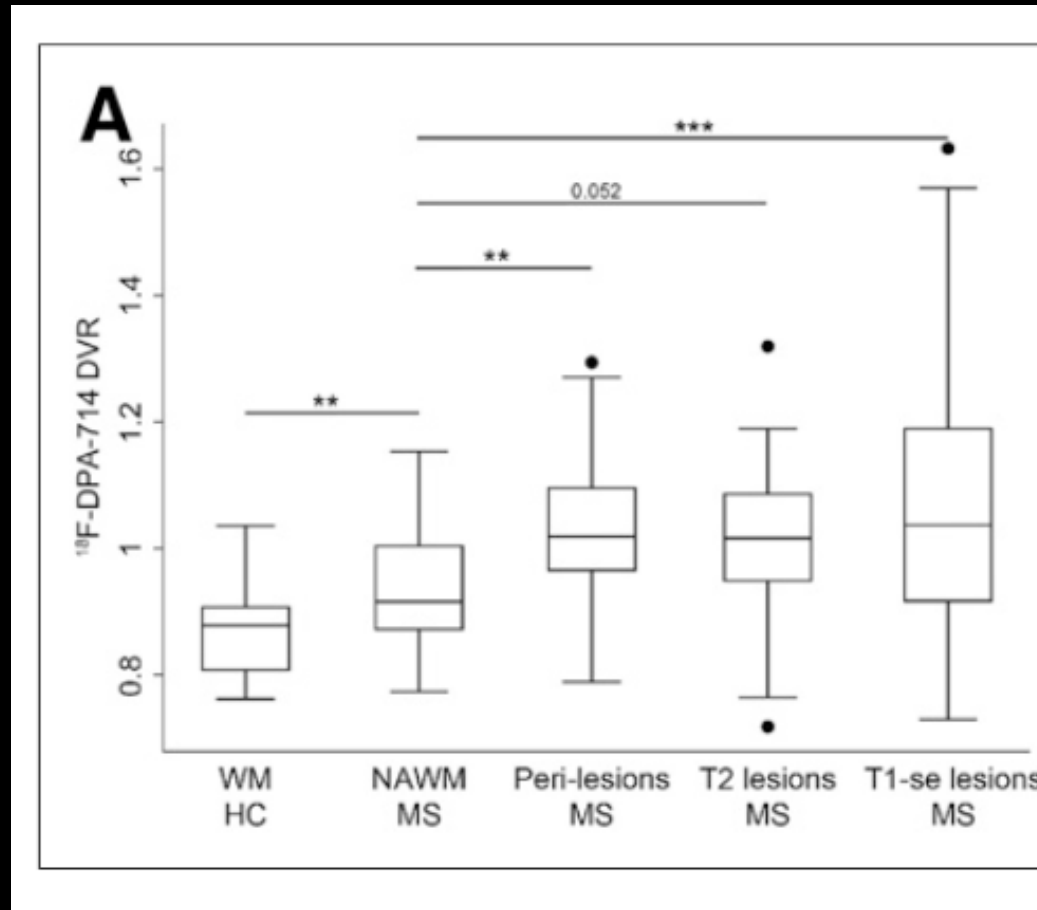
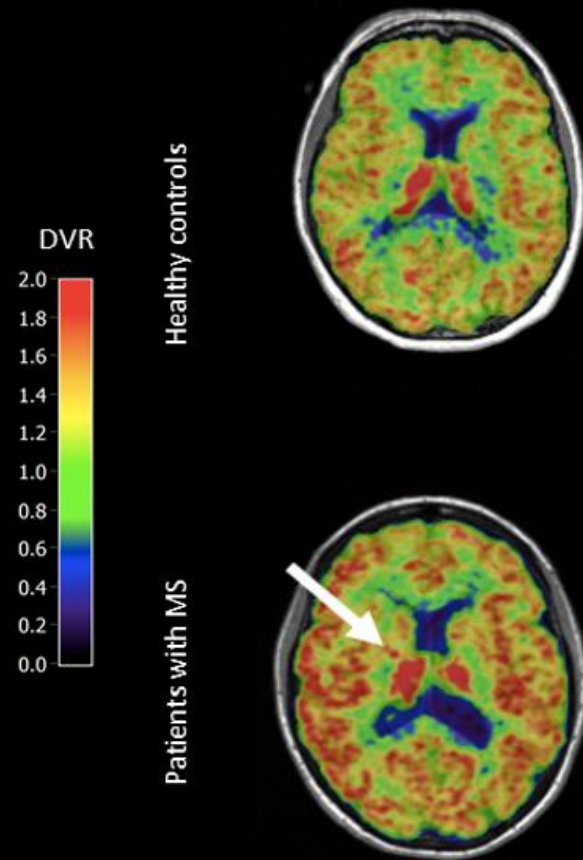
Correlation between
AIF based method and super DPA

- Automatic extraction of a reference region in the cortex using SuperDPA
- Extraction of the binding potential with a Logan reference tissue model

Improved reproducibility compared to cerebellar gray matter reference

- Why and how using PET to explore neuroinflammation in MS?
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TSPPO binding in the MS brain: region of interest analysis

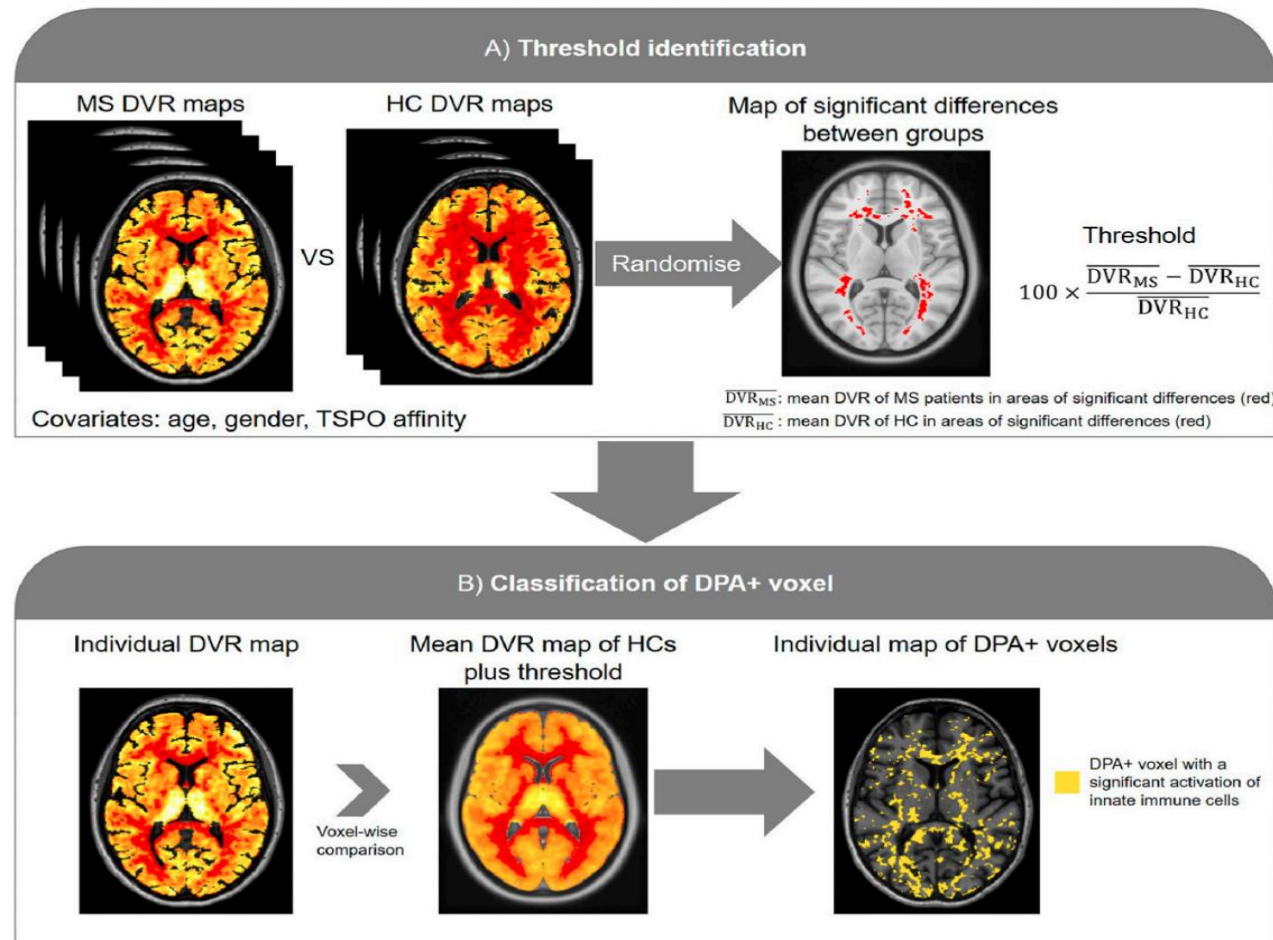


37 MS patients (12 RRMS, 14 SPMS, 11 PPMS) , 19 Healthy controls ; HRRT camera

Transversal analysis of neuroinflammation in key ROIs
Only non enhancing lesions analyzed

Individual mapping of innate immune cell activation with TSPO-PET

37 MS patients (12 RRMS, 14 SPMS, 11 PPMS) , 19 Healthy controls



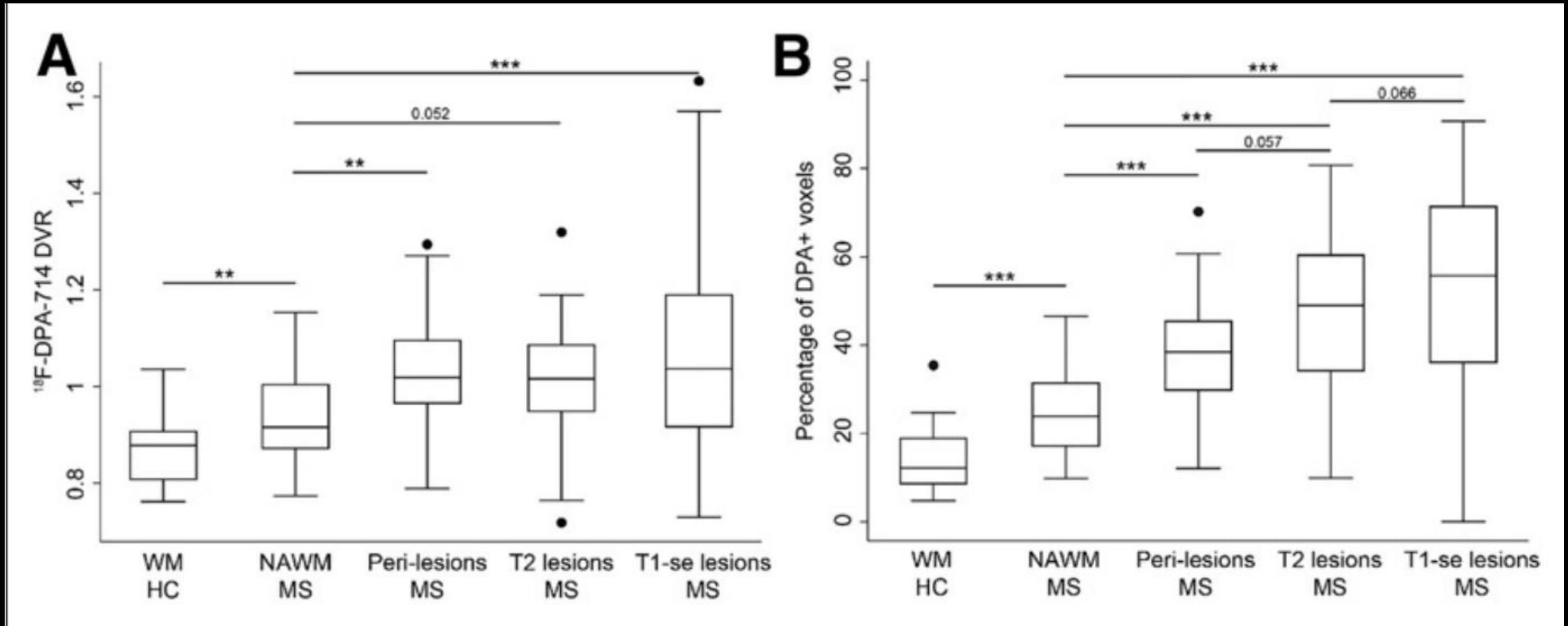
Goals :

- Minimise the influence of regional variation in unspecific binding and PVE
- Generate “comparable” maps in HAB and MAB
- Precise regional analysis at the lesional/sublesional level

Benoit et al, ECTRIMS/ACTRIMS 2017
Stankoff, ECTRIMS 2018
Bodini, Poirion et al, J Nuc Med 2020

Individual mapping of innate immune cell activation with TSPO-PET

37 MS patients (12 RRMS, 14 SPMS, 11 PPMS) , 19 Healthy controls

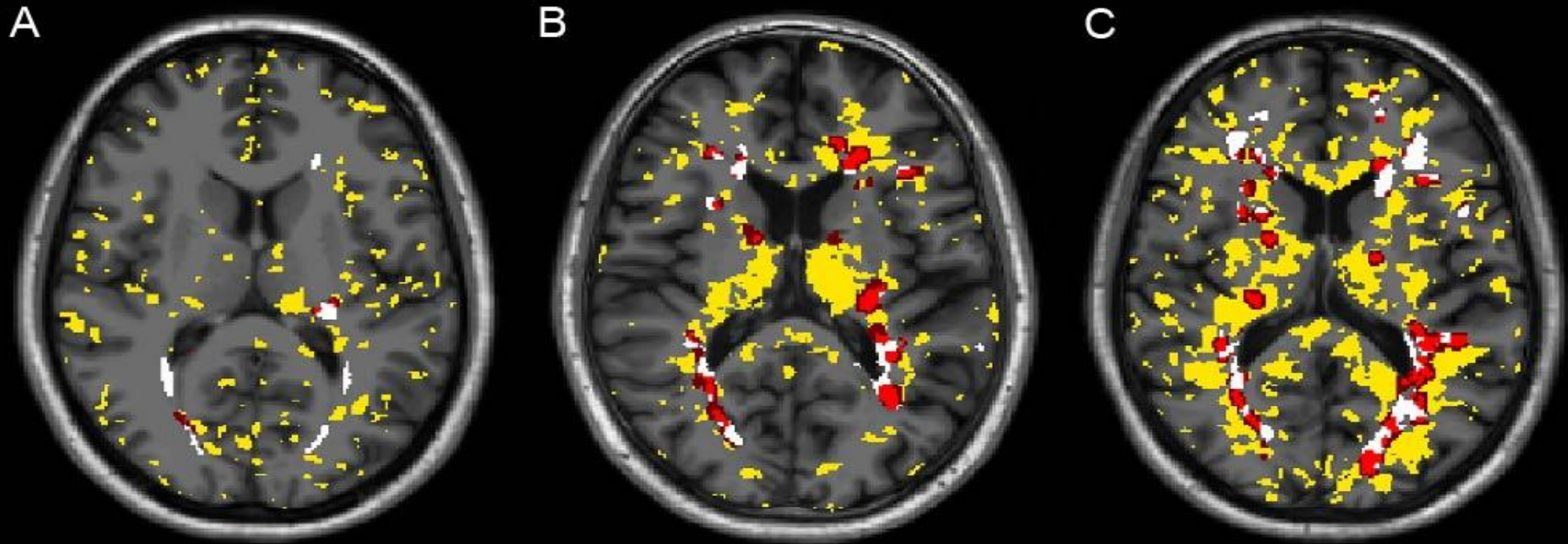


ROI analysis

Mapping of DPA active voxels

Only non enhancing lesions analyzed

Individual mapping of innate immune cell activation with TSPO-PET

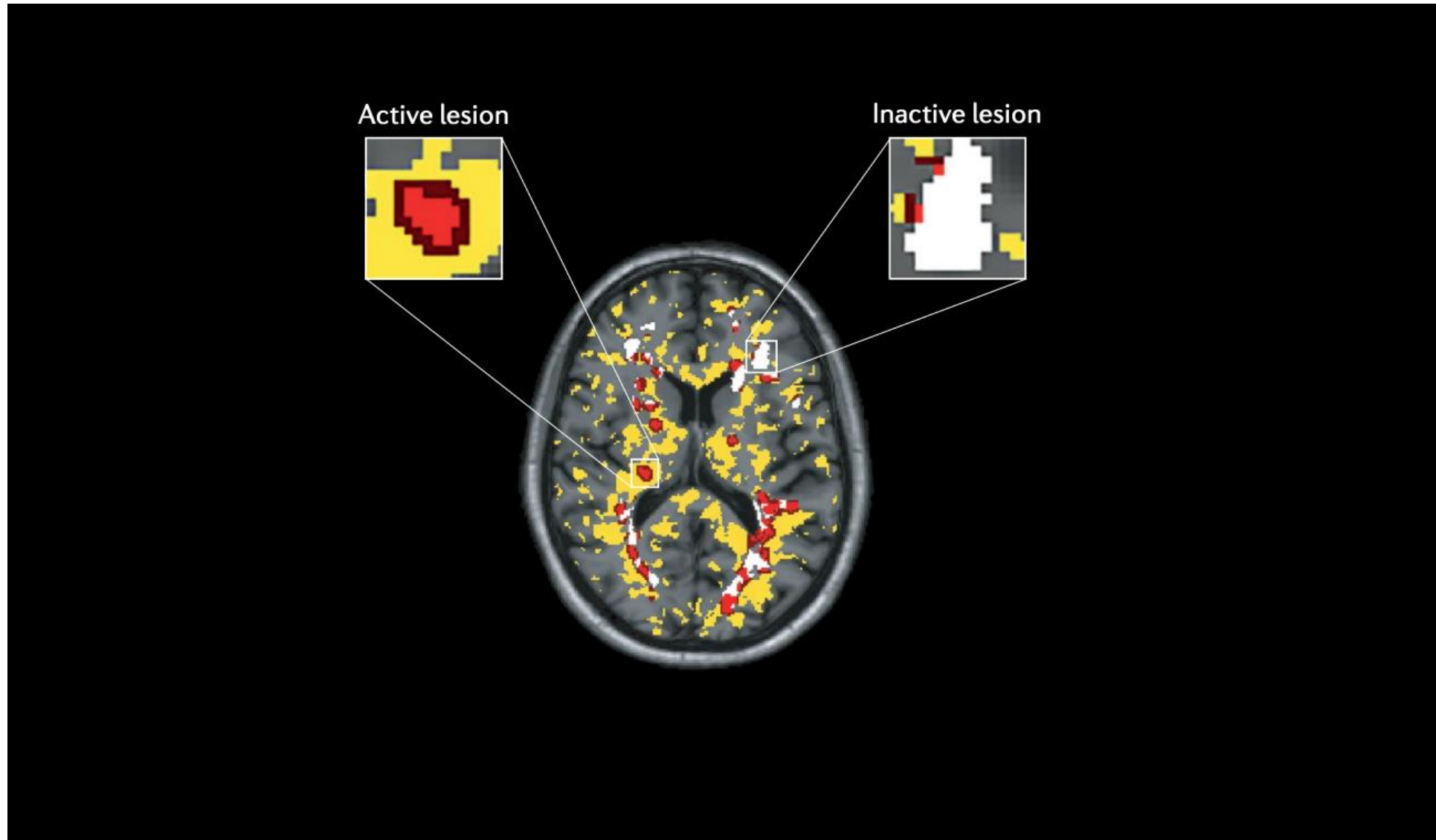


An innovative post-processing approach to generate individual maps of innate immune cells
Heterogeneous profiles of neuroinflammation depending on subjects
(Bodini, et al, J Nuc Med 2020; Nature Reviews Neurology 2021)

38 MS patients (13 RRMS, 14 SPMS, 11 PPMS) , 19 Healthy controls

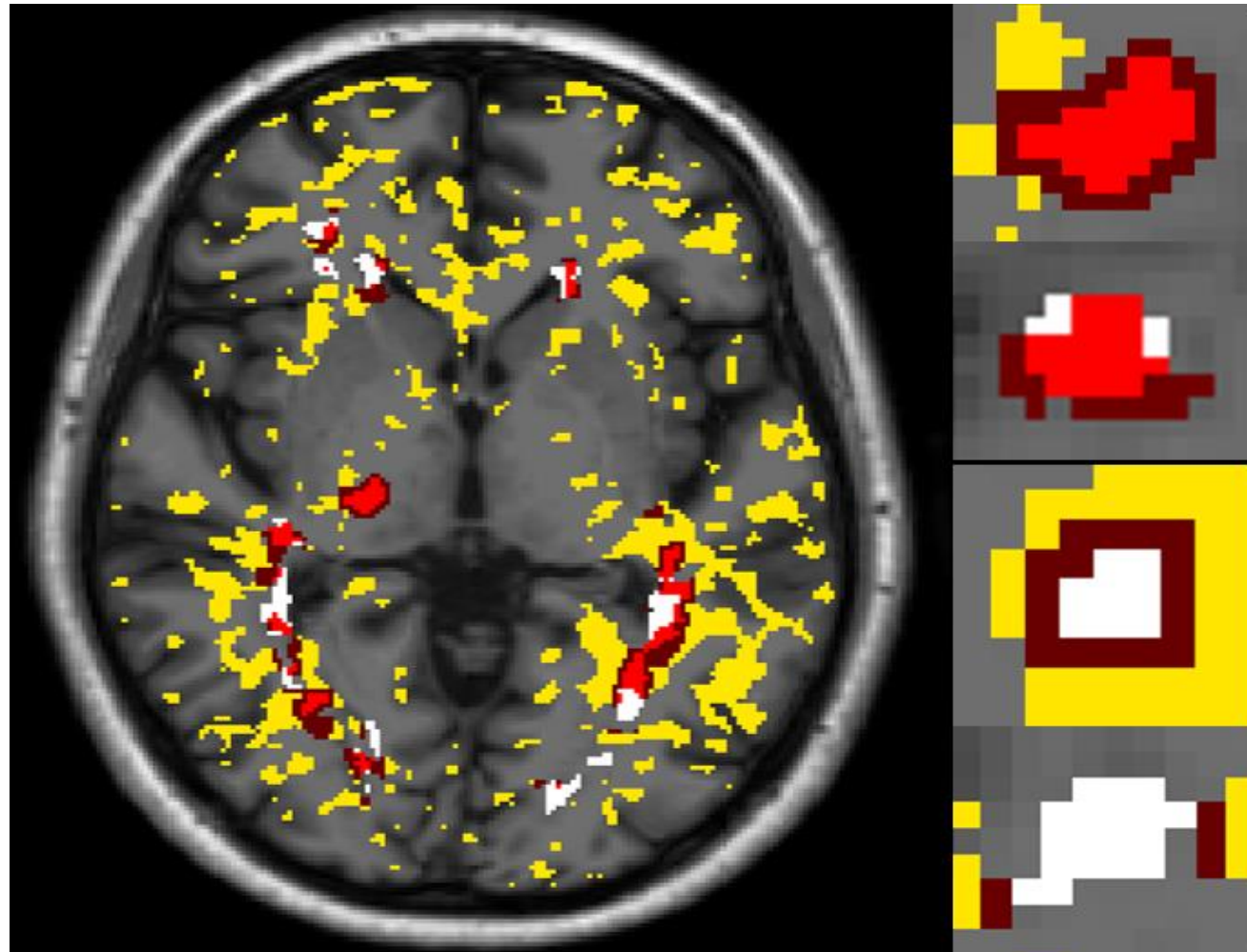
Individual mapping of innate immune cell activation with TSPO-PET

37 MS patients (12 RRMS, 14 SPMS, 11 PPMS) , 19 Healthy controls



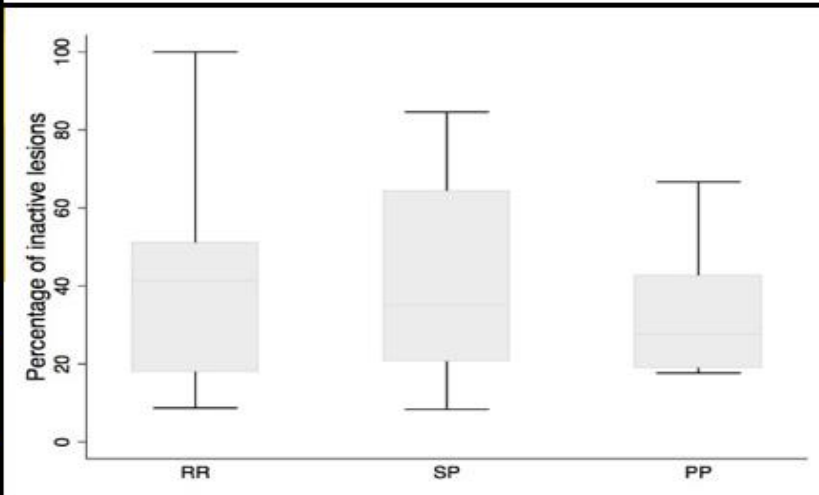
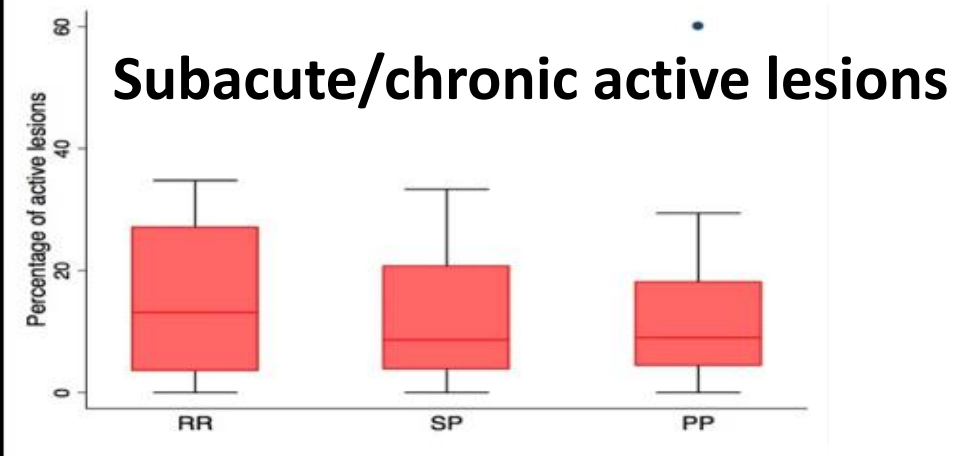
Benoit et al,ECTRIMS/ACTRIMS 2017; Stankoff et al, Brain Pathol, 2019; Bodini, Poirion et al, J Nuc Med, 2020

Active lesions identified using [^{18}F]DPA-714 PET



Classification of each lesion according to the innate immune cells content

> 50% activated voxels in individual lesions

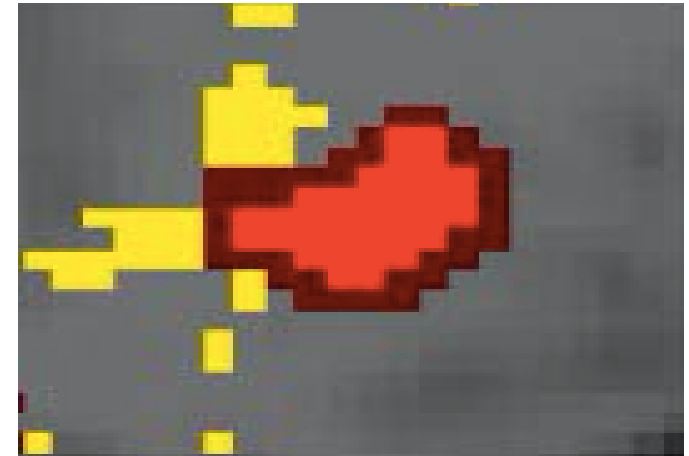


<10% activated voxel in lesions

Identifying hidden innate immune cell activation [18F]DPA-714

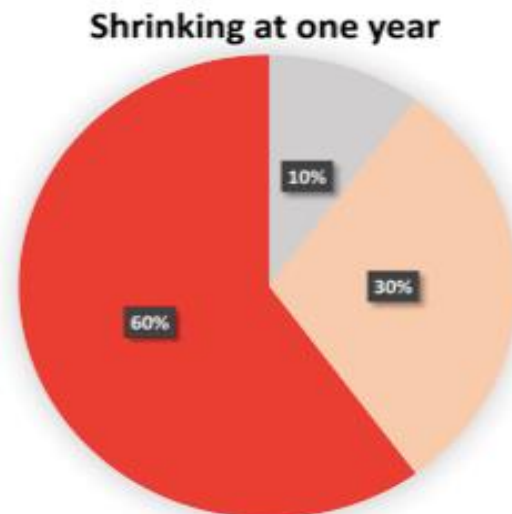
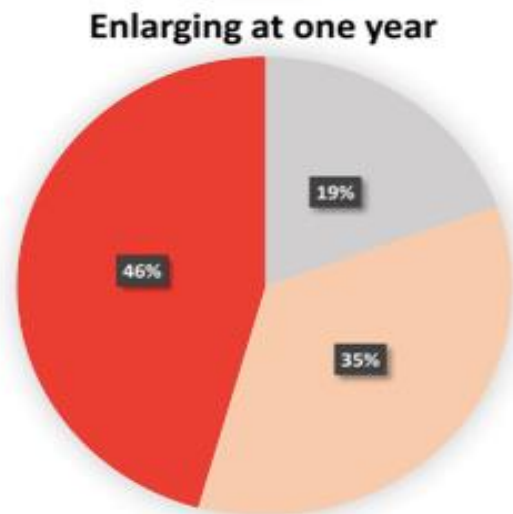
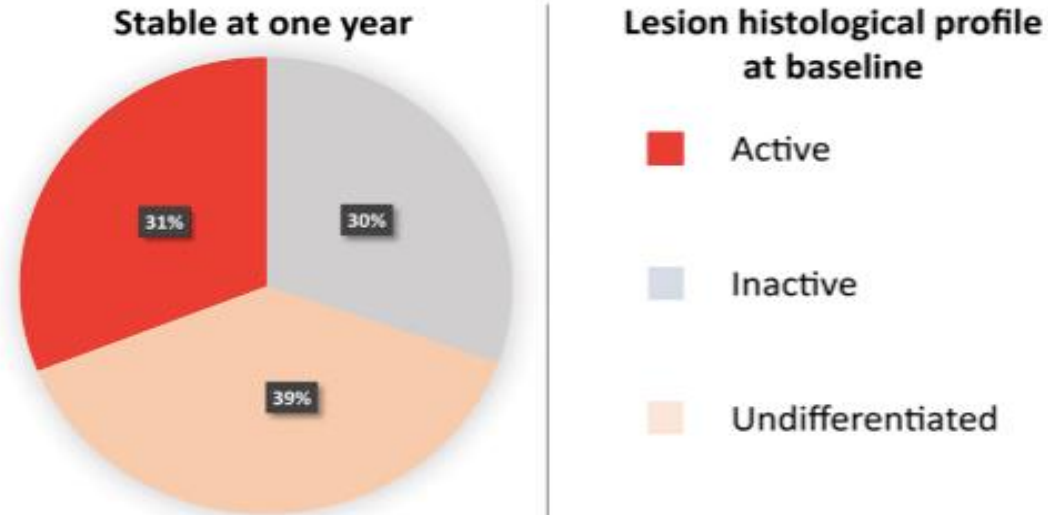


1.8% of total lesions classified as *active*
based on Gd+ T1 SE sequences



37.1% of total lesions were defined as
active based on DPA-PET

Dynamic evolution of [18F]DPA-714 subacute-chronic active lesions



10% of all lesions changed volume after one year (7% enlarged, 3% shrank).

Active lesions were more likely to enlarge or shrink over time

Subacute and chronic active lesions and trajectories of clinical disability

N. OF ACTIVE LESIONS

Beta-coeff=0.76

p=0.001

EDSS step change

**EDSS step change during the 2 years
preceeding study entry**

**Innate immune cells activation:
a marker of individual trajectory**

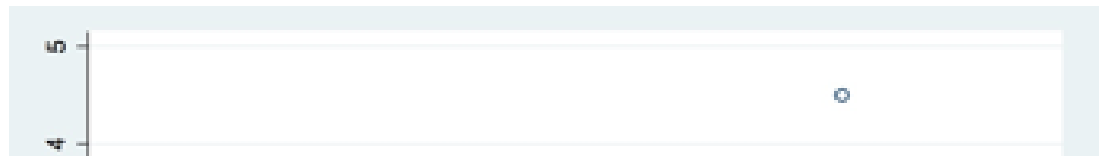
of disability worsening

% ACTIVATION in the Perilesions

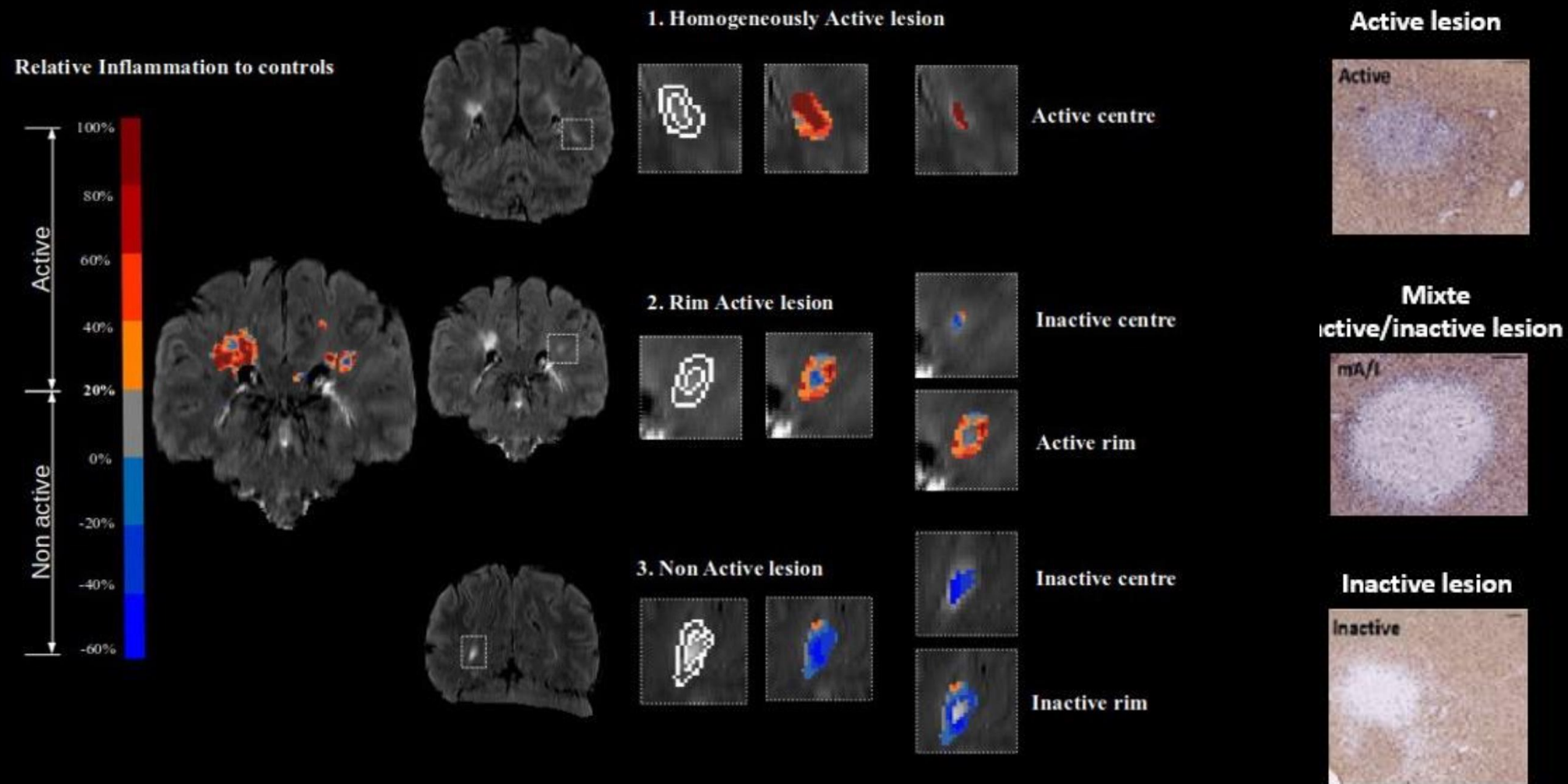
Beta-coeff=0.50

p=0.01

EDSS step change



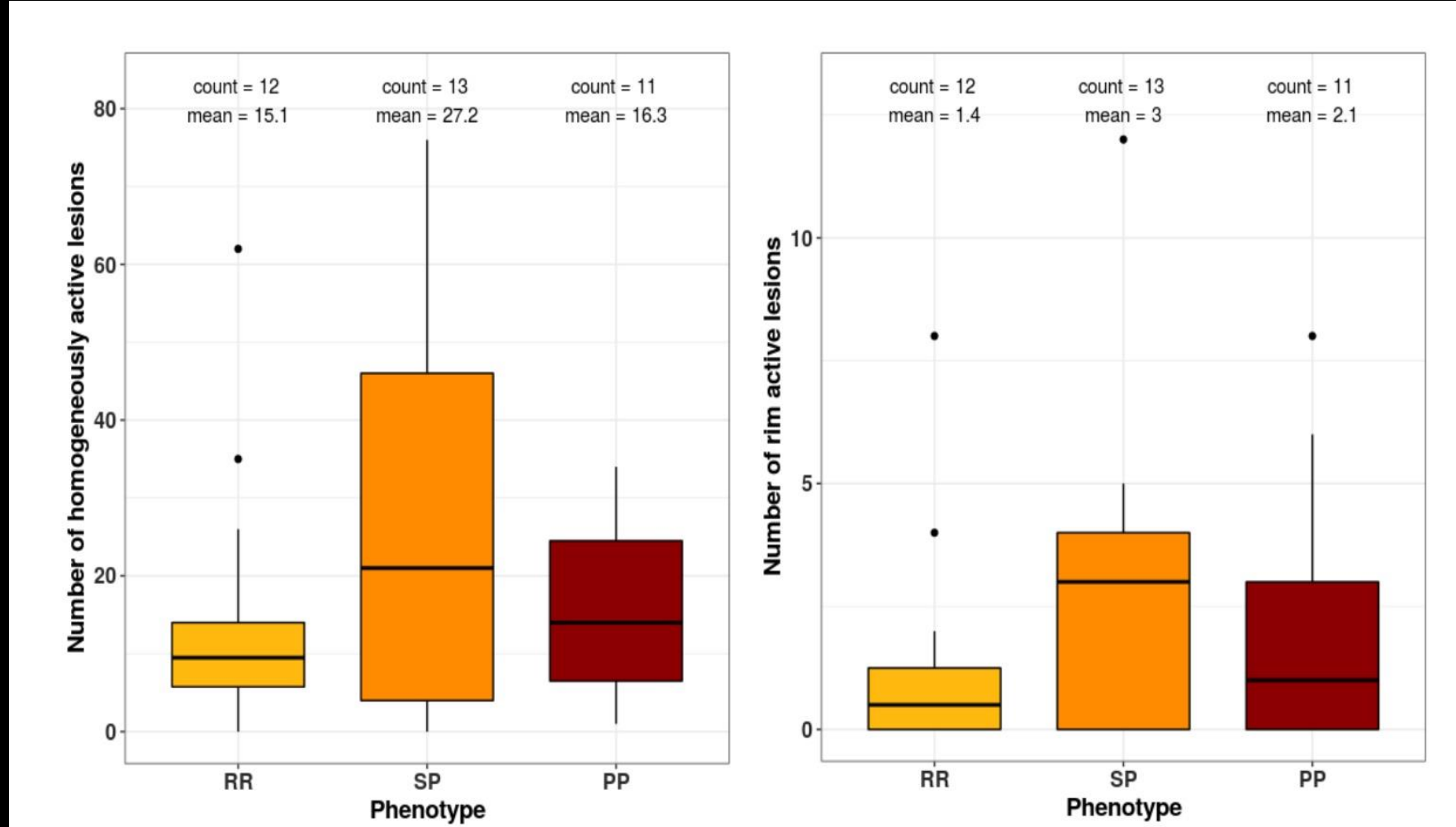
Exploring the smouldering component of MS lesions: a new PET-based classification



Homogeneously active (53%) and mixte active/inactive lesions (6%)

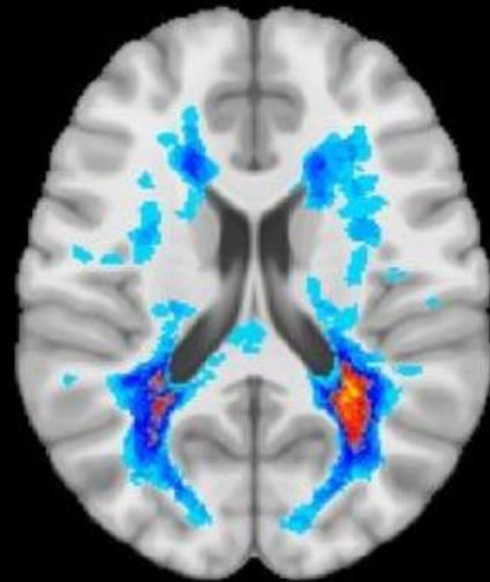
Hamzaoui, Garcia et al 2022

Exploring the smouldering component of MS lesions: a new PET-based classification

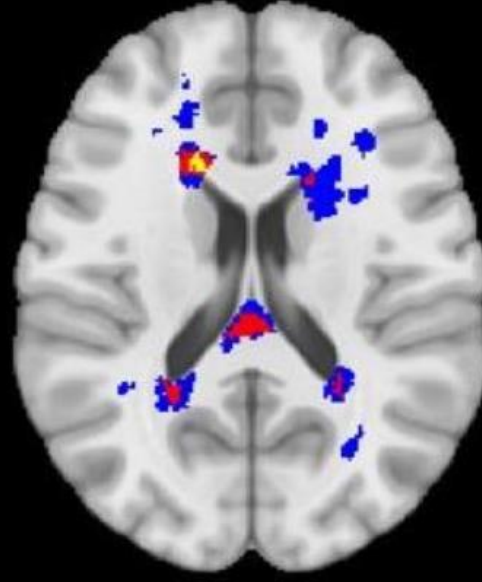


Homogeneously active and rim-active lesions at all MS stages
Bodini, Poirion et al, 2020; Hamzaoui, Garcia et al 2022

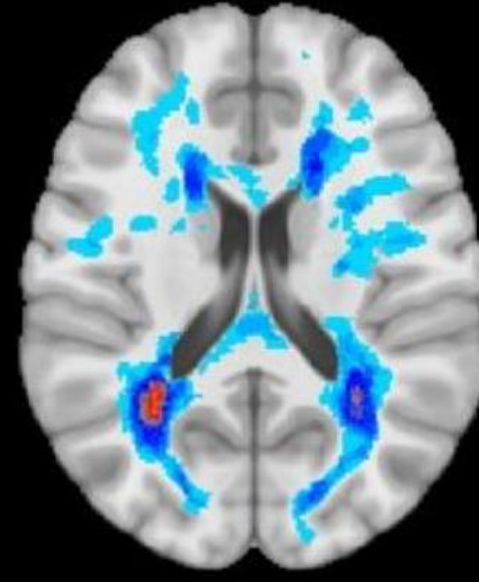
The regional distribution of lesions according to their neuroinflammatory profiles



Lesion probability map for homogeneously active lesions



Lesion probability map for rim active lesions



Lesion probability map for non active lesions

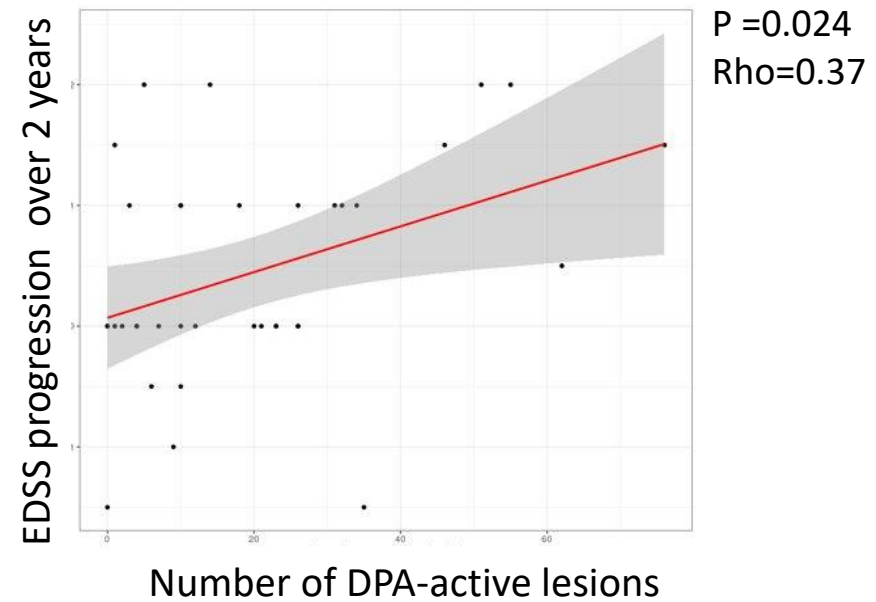
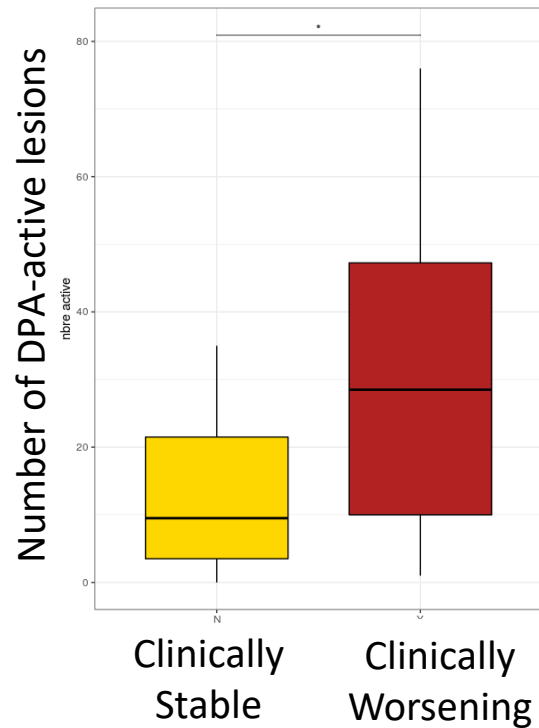
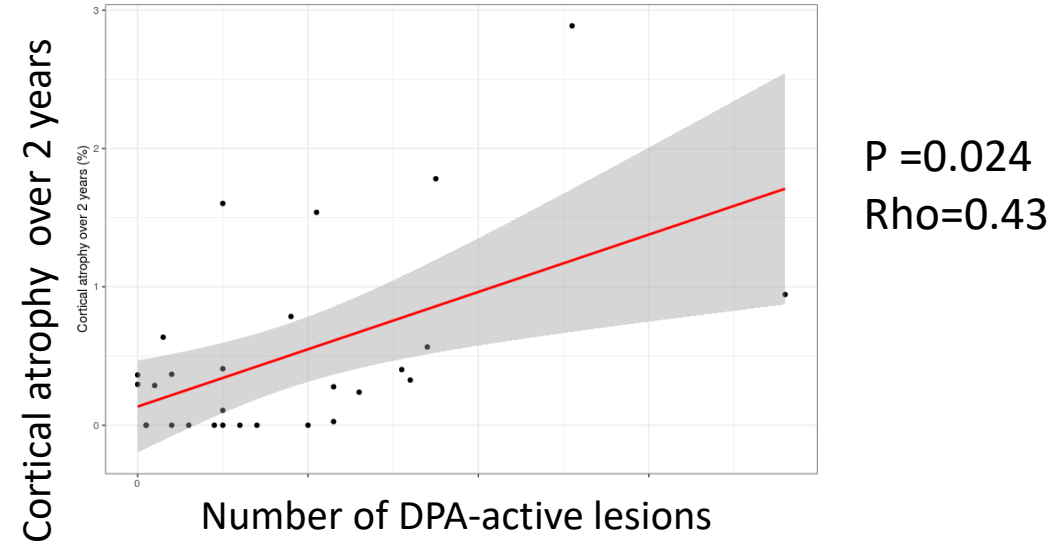
Probability of a voxel to be in a lesion



High probability

Low probability

Homogeneously active lesions predict cortical atrophy and clinical progression



- Why using PET to explore neuroinflammation in MS?
- Exploring neuroinflammation in lesions and normal-appearing tissues in MS
- **Regionalization of neuroinflammation in the MS brain**
- A dysfunction of the Brain/CSF barrier at the choroid plexus level

Is chronic neuroinflammation the result of CSF/meningeal-derived factors?

A periventricular gradient of MTR

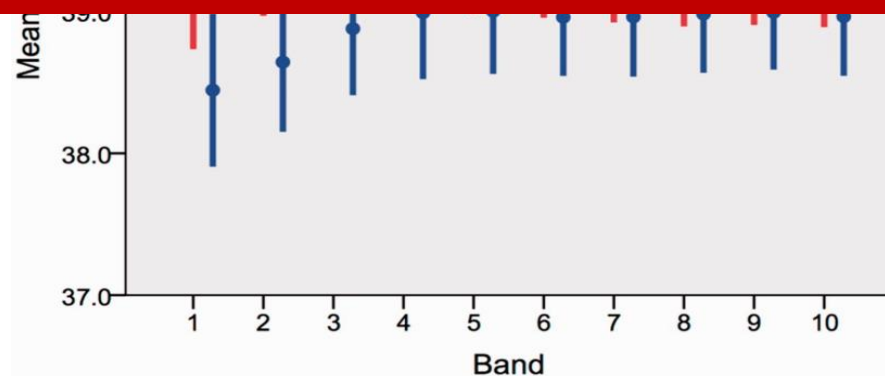


Relationship between cortical and periventricular NAWM damage

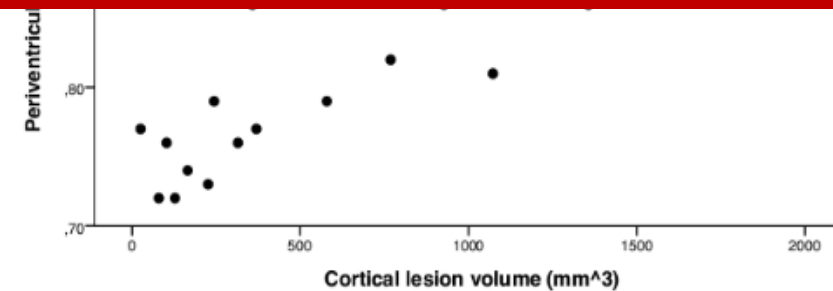


Is the gradient of tissue damage linked with innate immune cell activation?

What is the clinical relevance?



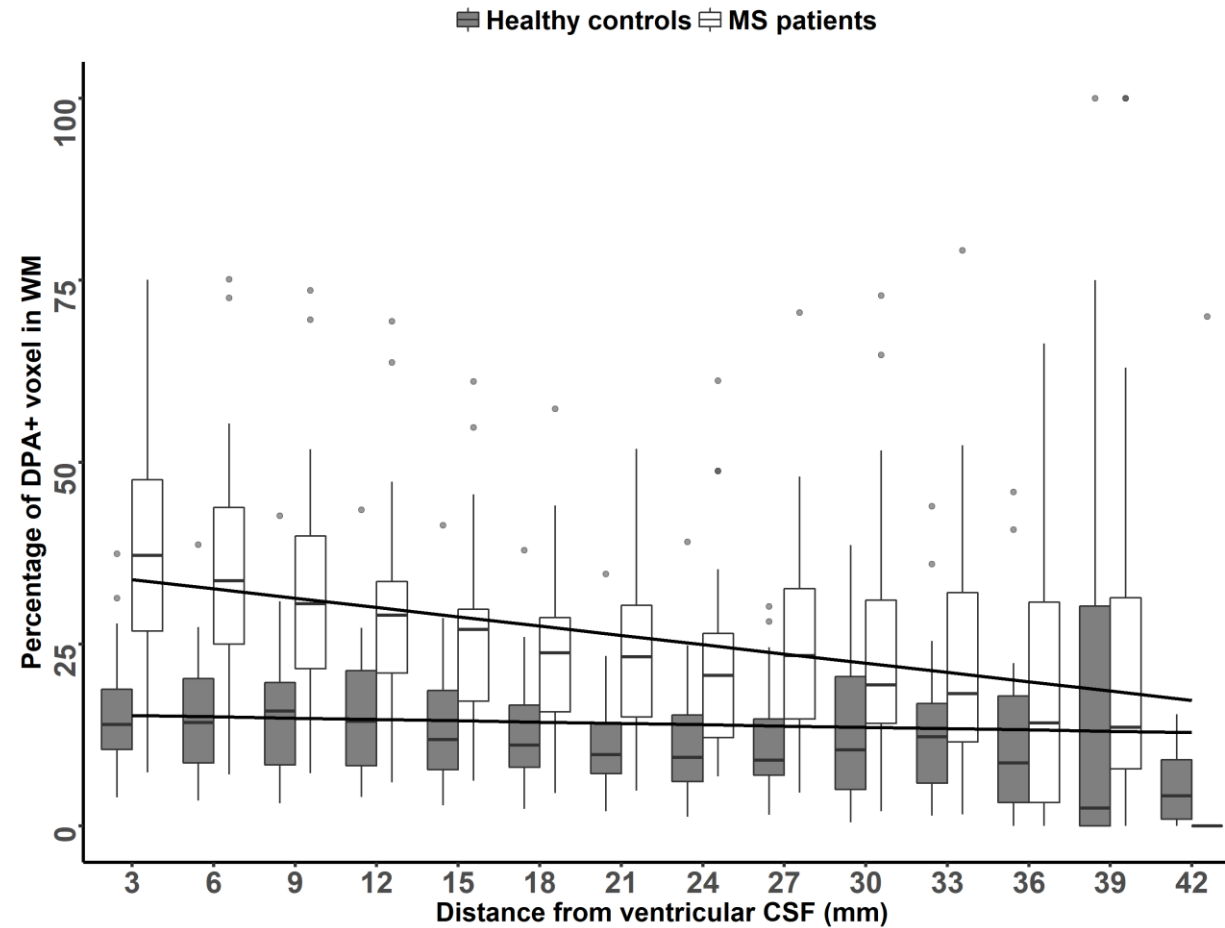
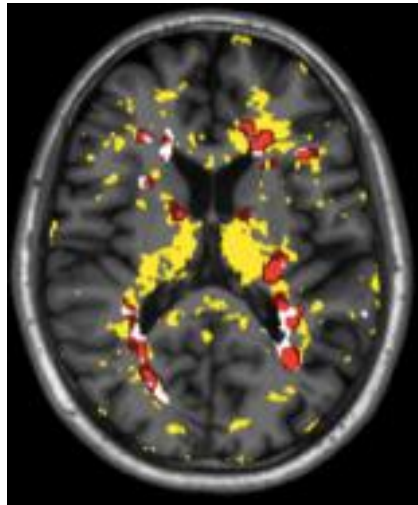
Liu et al, Brain 2015



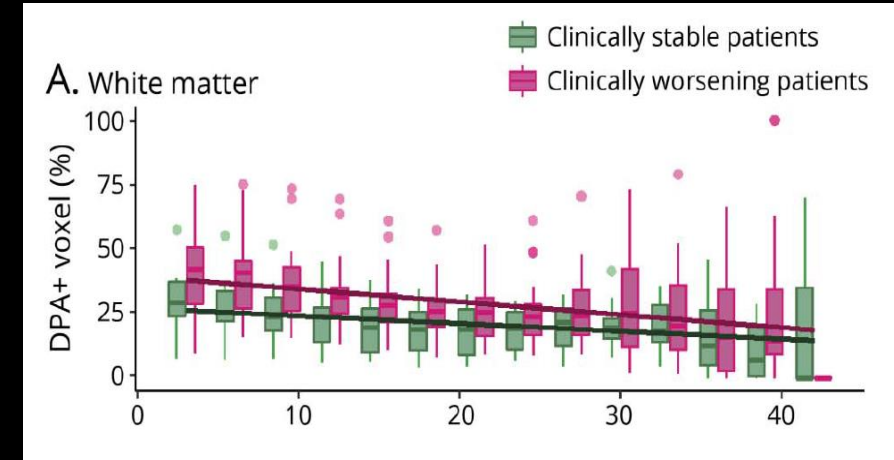
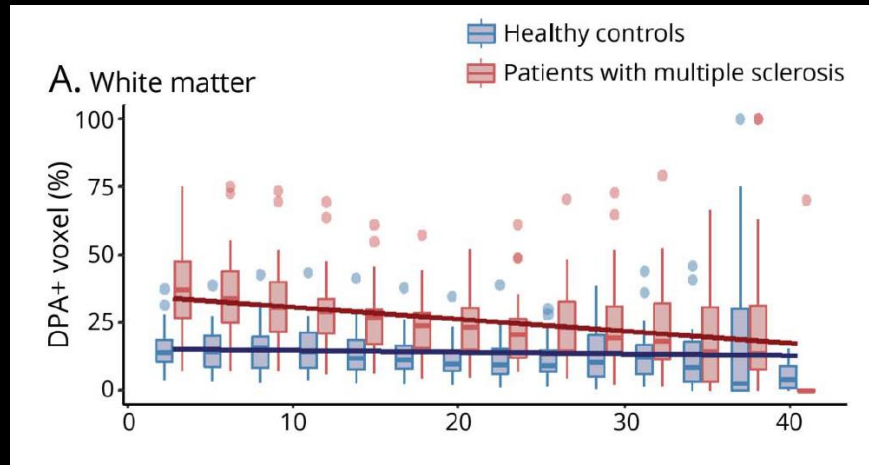
Pardini et al, NeuroImage Clin 2017

A periventricular gradient of innate immune cell activation measured with PET

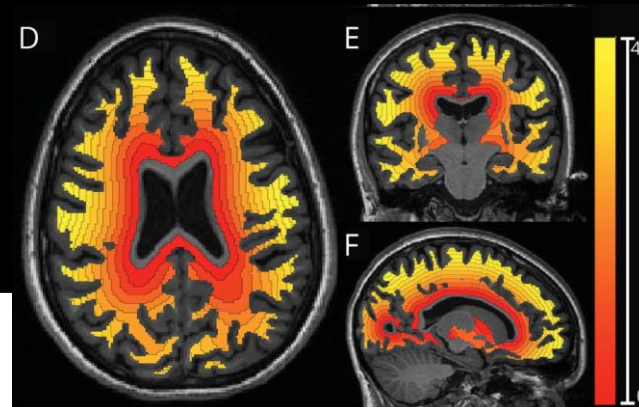
White matter



Periventricular innate immune cell activation correlates with disability progression

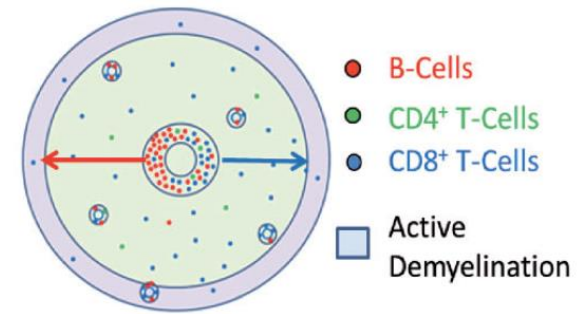


- Innate immune cell activation in MS predominates in periventricular regions
- Activation of microglia and macrophages in periventricular normal-appearing white matter is associated with microstructural damage and disability worsening



A periventricular gradient of innate immune cell activation: the hypotheses

- Compartmentalized inflammation in the perivenular space predominates around ventricles?



Machado-Santos, Brain, 2018

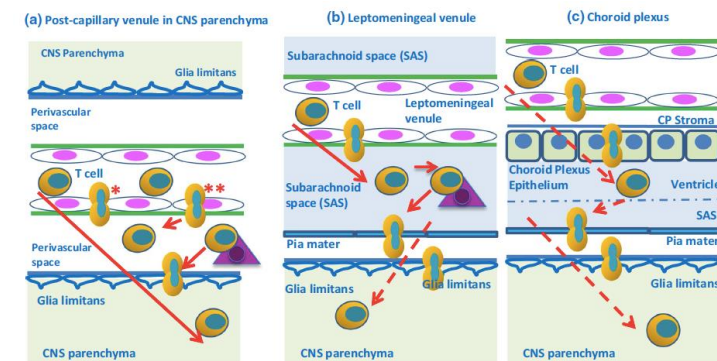
- CSF derived pro-inflammatory factors?

Cytokines, chemokines (Magliozzi, Ann Neurol 2018);

Fibrinogen (Peterson et al, Nat Rev 2018);

Ceramide (Wentling, PNAS, 2019) ...

- The blood-CSF barrier (choroid plexus) ?



Engelhardt et al, Acta Neuropathol, 2016

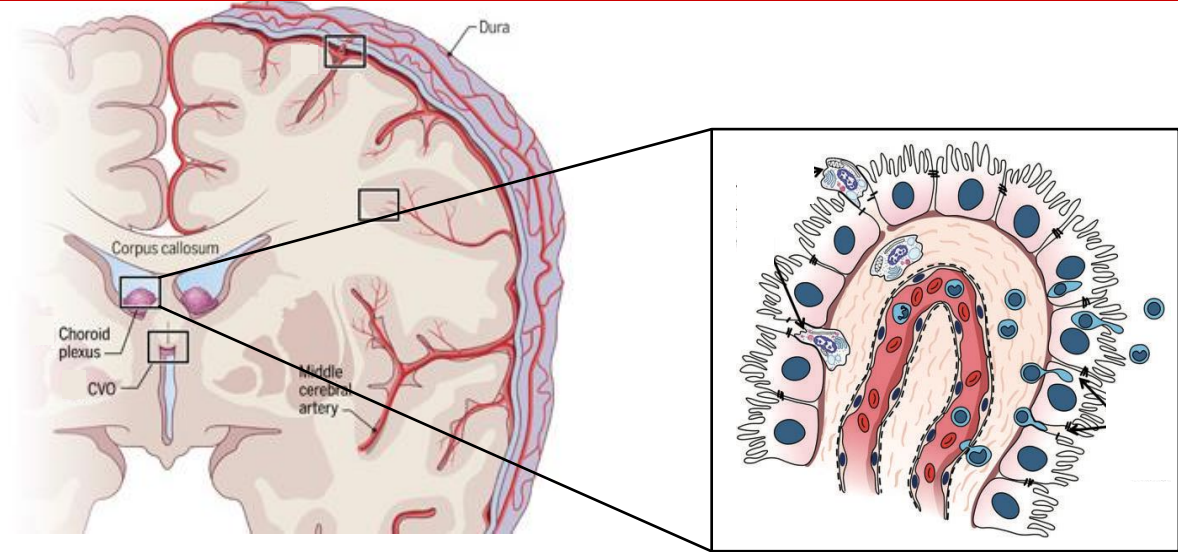
- Why using PET to explore neuroinflammation in MS?
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- **A dysfunction of the Brain/CSF barrier at the choroid plexus level**

Choroid plexuses in animal models of MS and in post-mortem brains

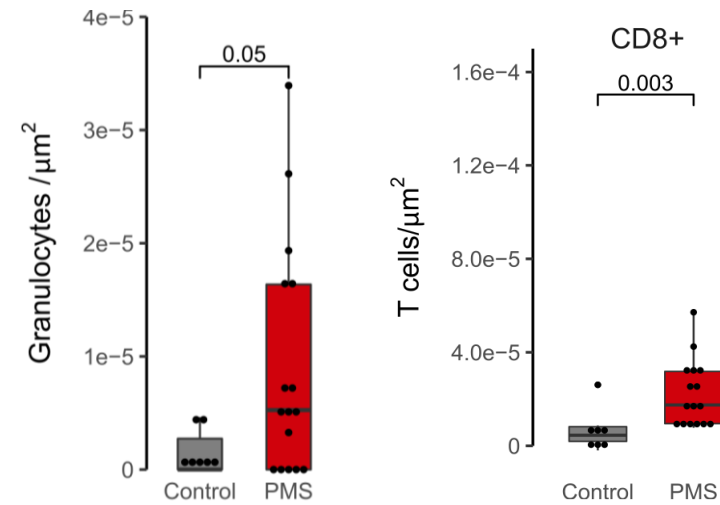
The CPs act as a **unique neuro-immunological interface** integrating CNS signals with those from circulating immune cells.

In the EAE model of MS, the CPs allow the initial entry of encephalitogenic T-Helper cells into the CNS.

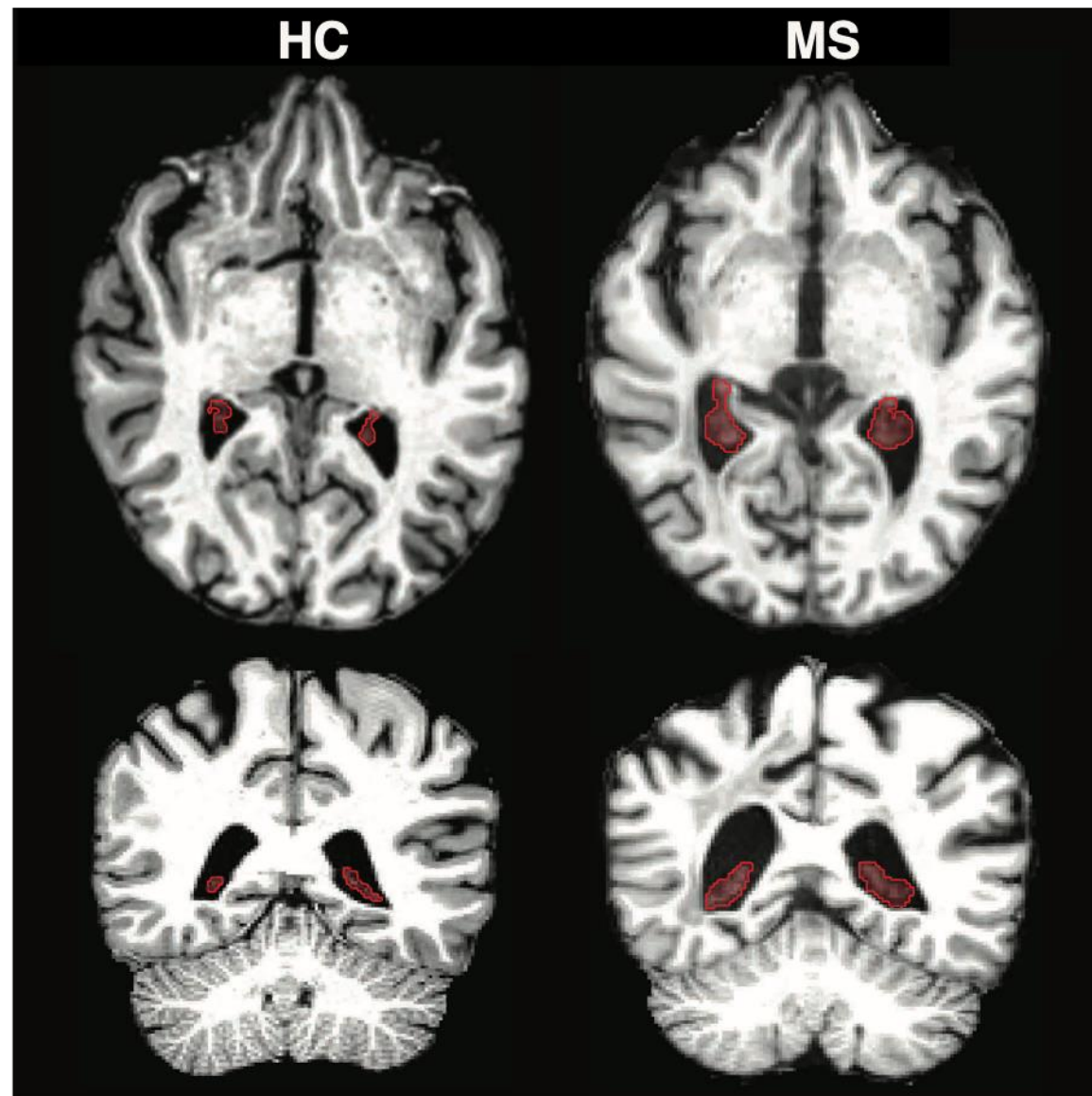
Post-mortem studies have shown **infiltrating innate cells, granulocytes** and **CD8+ T cells** in the CPs of MS.



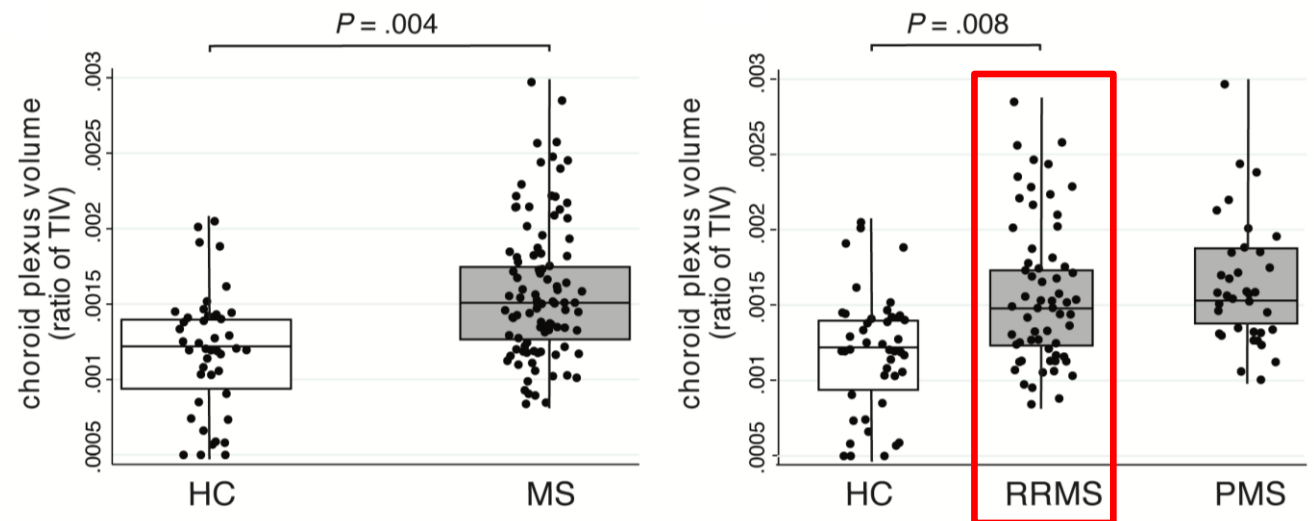
Kaur C et al, 2016



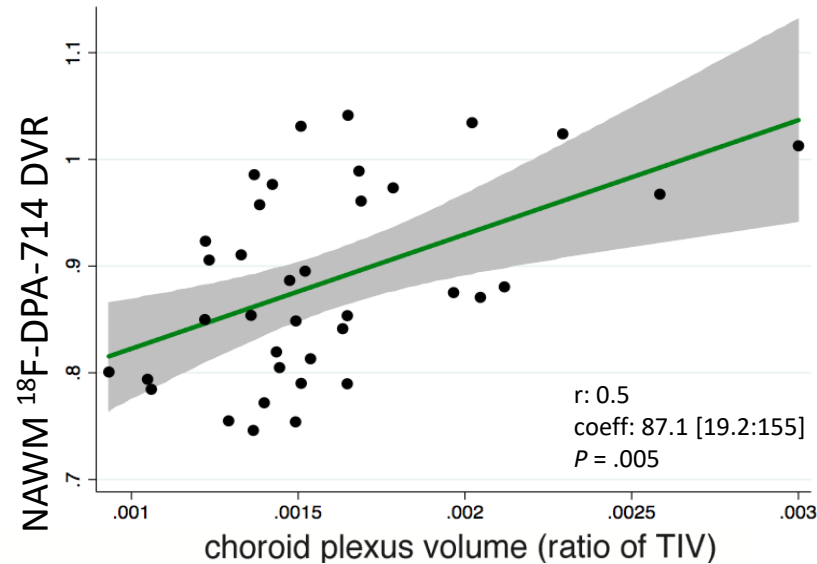
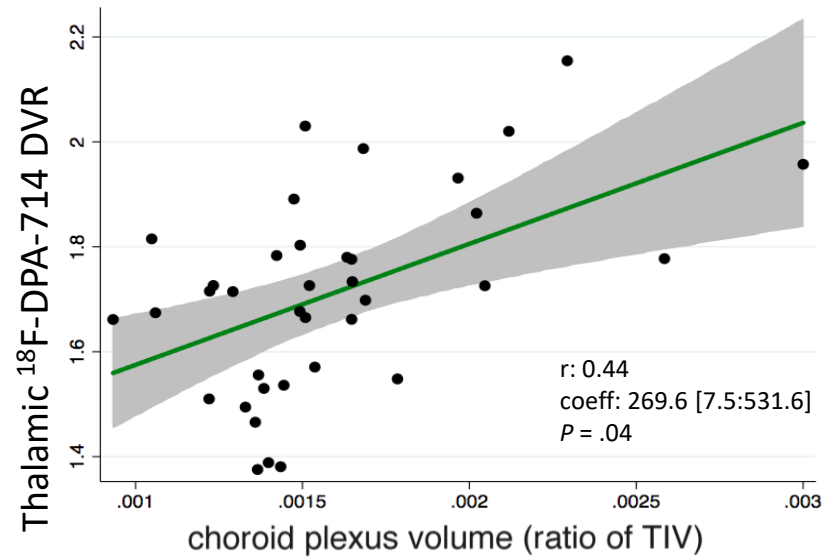
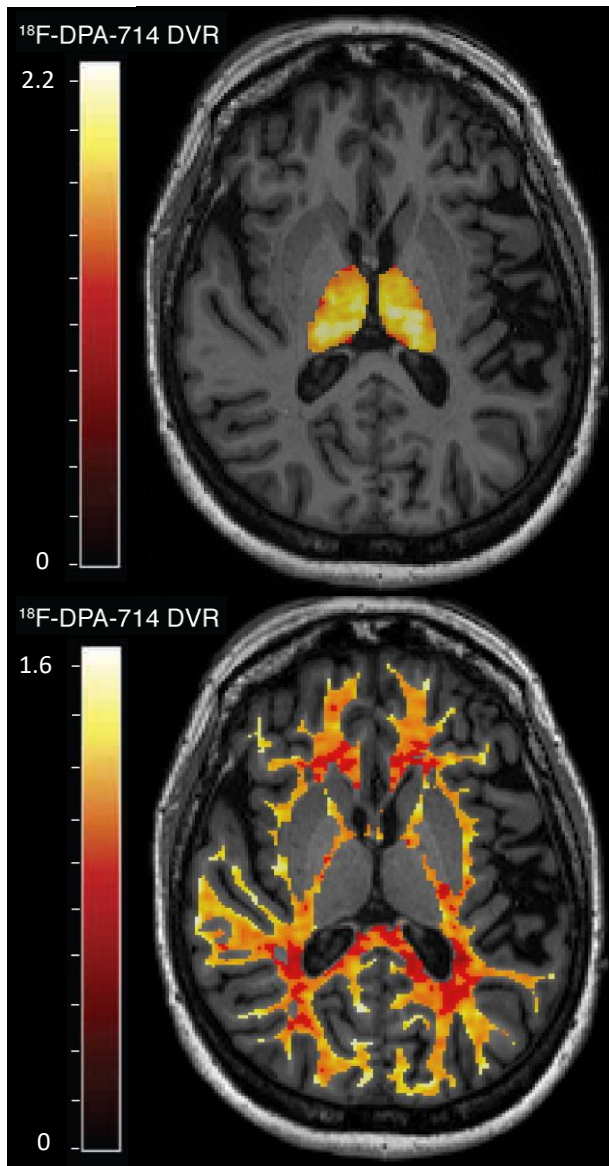
Choroid plexuses in people with MS are larger than in healthy controls



Choroid plexus volume is **35% greater** in patients with MS compared with HC

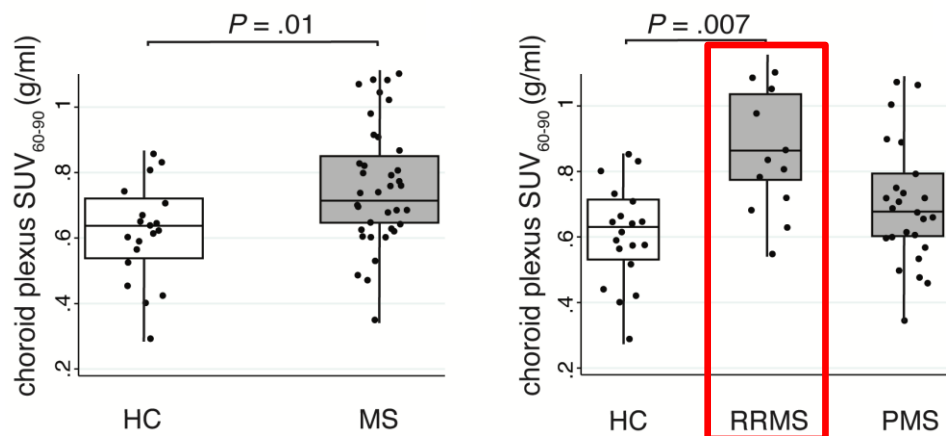
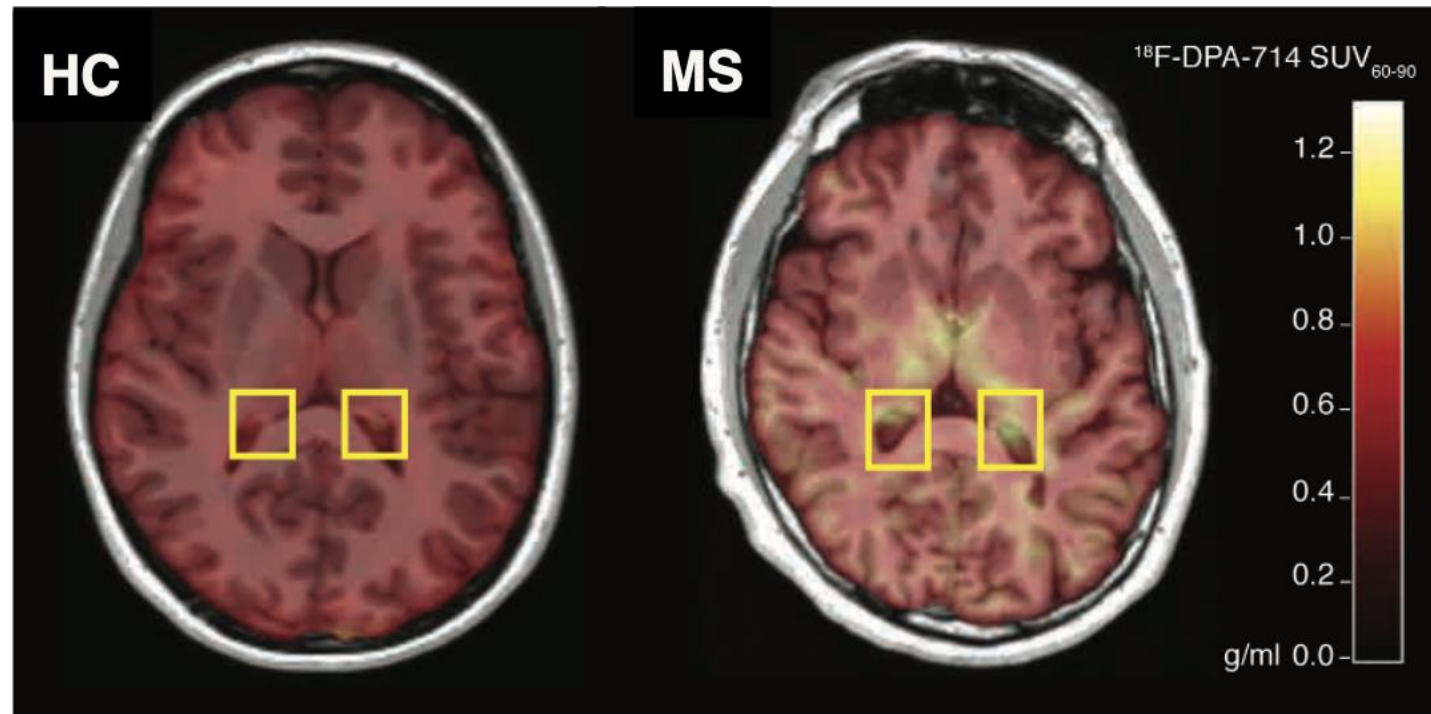


CP volume correlates with parenchymal neuroinflammation

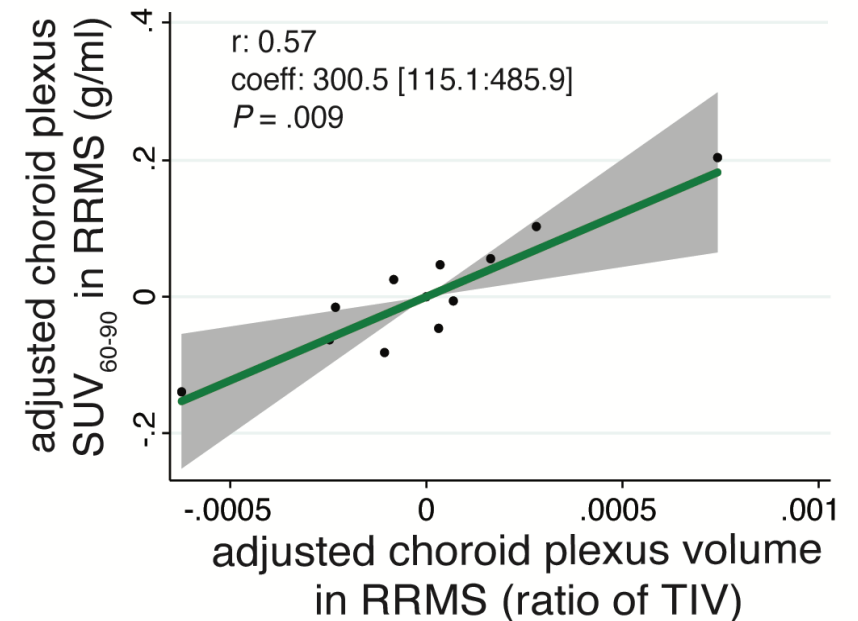


Larger CPs are associated with
**greater innate immune cell activation in the
thalami and the NAWM**
as measured by ¹⁸F-DPA-714 DVR

TSPO PET reveals inflamed choroid plexuses in MS

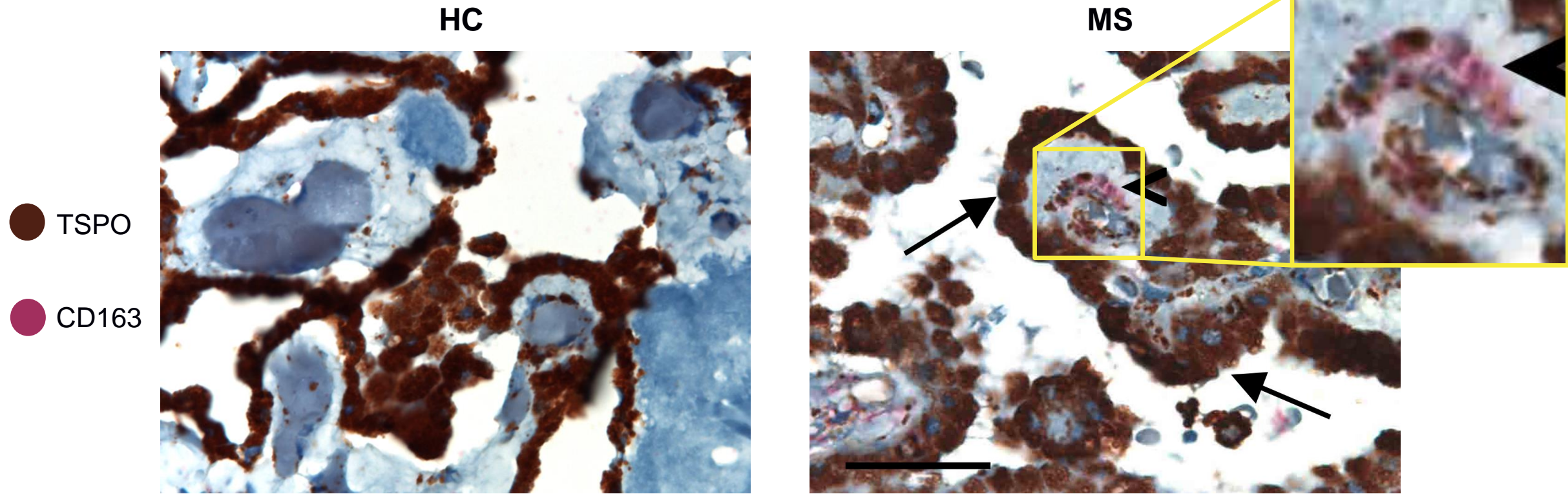


Choroid plexuses in MS have a **18.5% higher DPA uptake** compared with HC



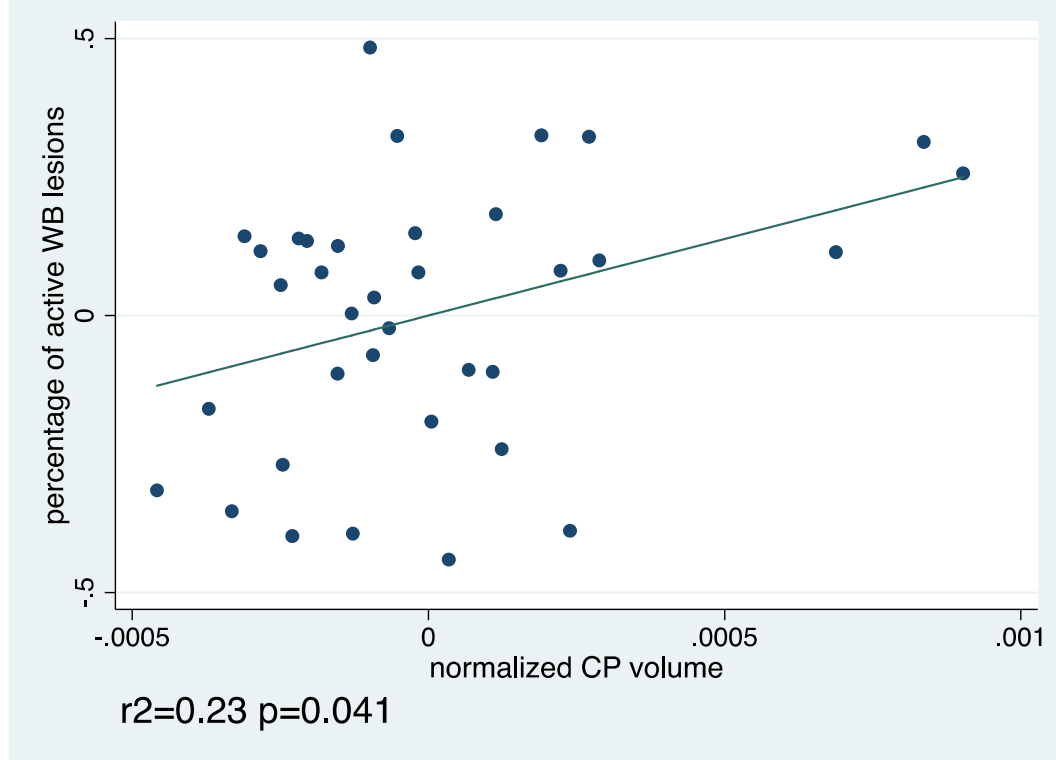
Ricigliano et al, Radiology 2021

[18F]DPA-714 uptake in MS Choroid plexuses

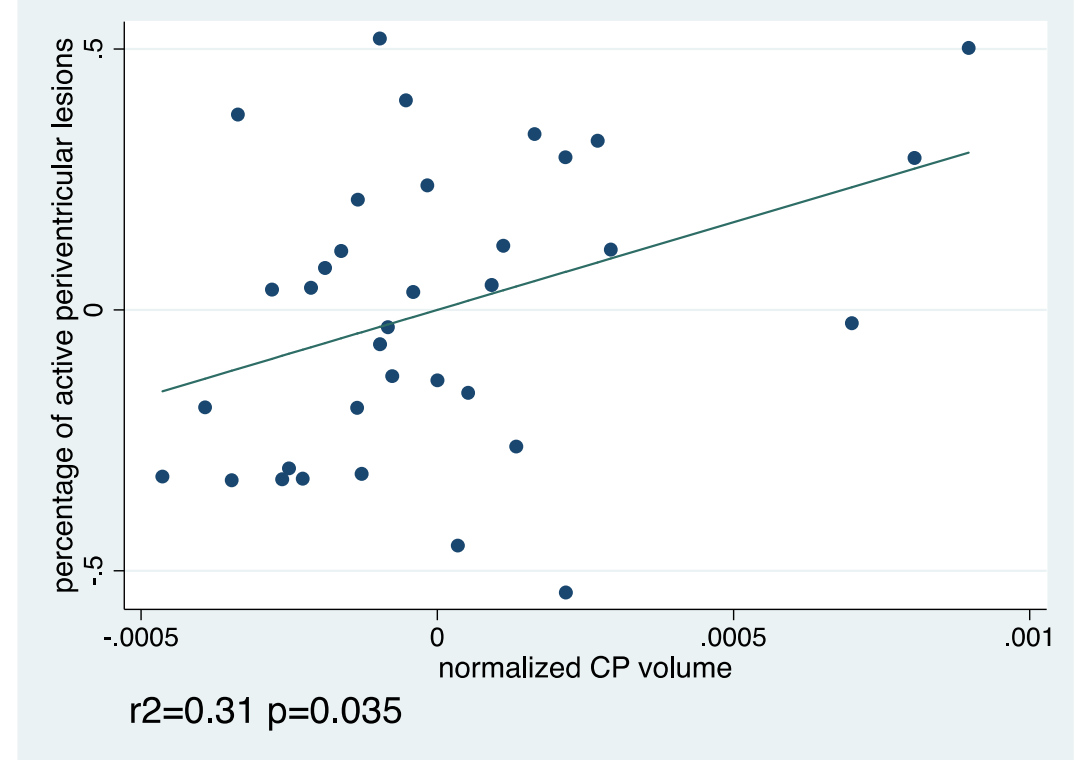


Infiltrating **CD163+** macrophages contribute to the higher TSPO expression in MS CPs, in addition to TSPO expression in the CP epithelium of both MS and HC

Relationship between choroid plexus volume and DPA-active lesions



RRMS: $r^2=0.52$ $p=0.011$



RRMS: $r^2=0.74$ $p<0.001$

larger CPs



higher percentage of active lesions in the whole brain, particularly in the periventricular region

Conclusions

- TSPO PET is applicable in clinical studies without arterial sampling
- TSPO PET allows to generate regional individual mapping of neuroinflammation, reflecting innate immune cells density, in the MS brain
- Molecular imaging reveals smouldering component in the majority of lesions, that may start at the relapsing stage, and is linked to disability progression
- The smouldering component is regionalized in the brain and may involve a BCSFB dysfunction at the choroid plexus level

Acknowledgements

B Stankoff/ C Lubetzki ICM team

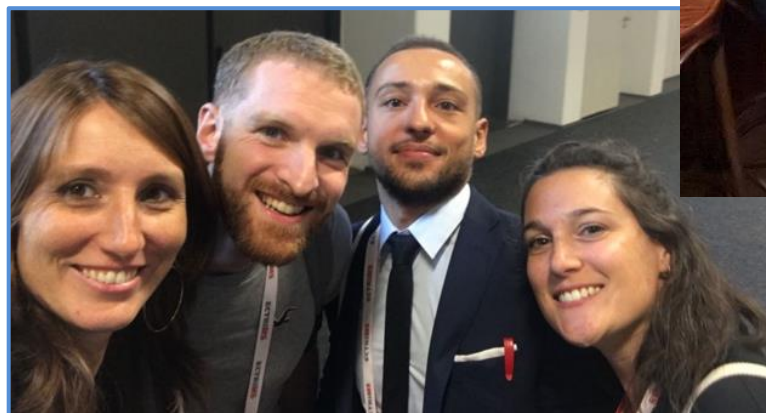
Repair in MS: from biology to clinical translation

Bruno Stankoff
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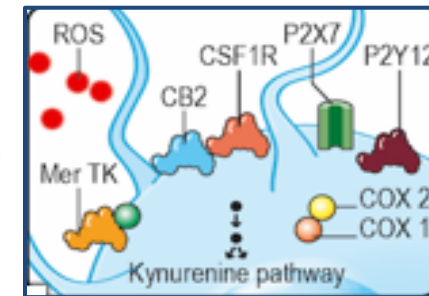
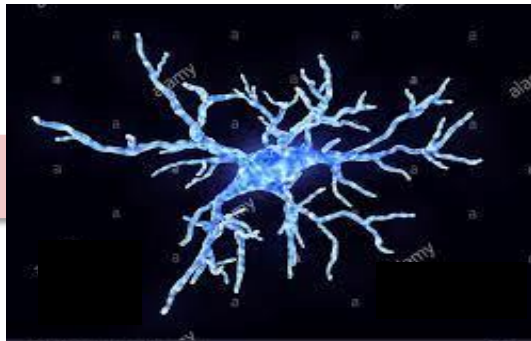
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Bertrand Kuhnast
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Novel perspectives in innate immune cell imaging



FUTURE TARGETS FOR MICROGLIA AND MACROPHAGES

Macrophage colony stimulating factor 1 receptor

PET imaging of microglia by targeting macrophage colony-stimulating factor 1 receptor (CSF1R)

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P2Y12R
P2X7R

The P2X₇ receptor tracer [¹¹C]SMW139 as an *in vivo* marker of neuroinflammation in multiple sclerosis: a first-in man study

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